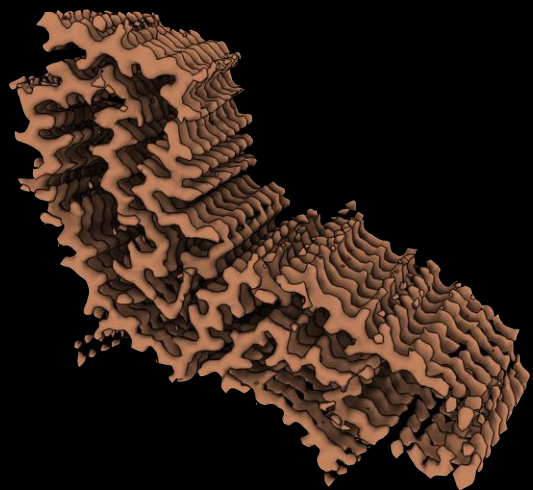


The image displays several cryo-electron tomograms of neurons, arranged in a grid-like pattern. Each tomogram shows a 3D reconstruction of a neuron, with various organelles and structures visible. The neurons are stained with green, blue, and red dyes, highlighting different components. The background is a dark, textured surface with a grid of small, light-colored dots, likely representing the cryo-EM grid. The text is overlaid on the top-left portion of the image.

Cryo-correlative imaging of prion strains
in neurons

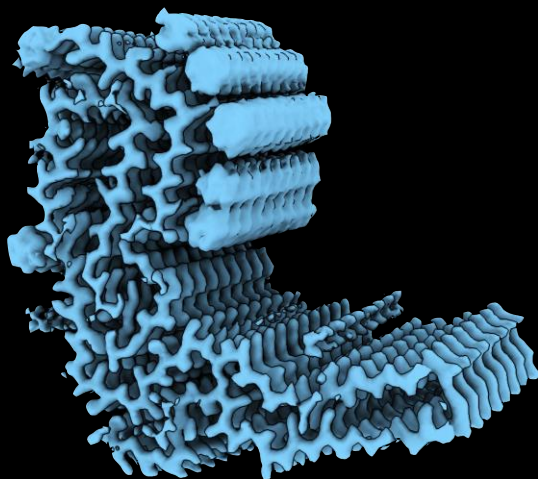
Thomas Trainer

263K



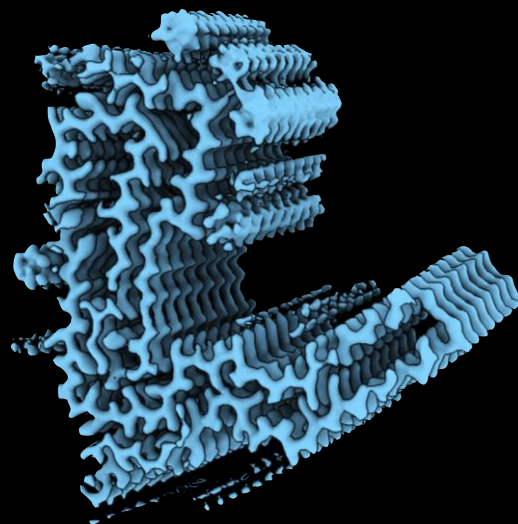
Kraus et al. 2021

RML



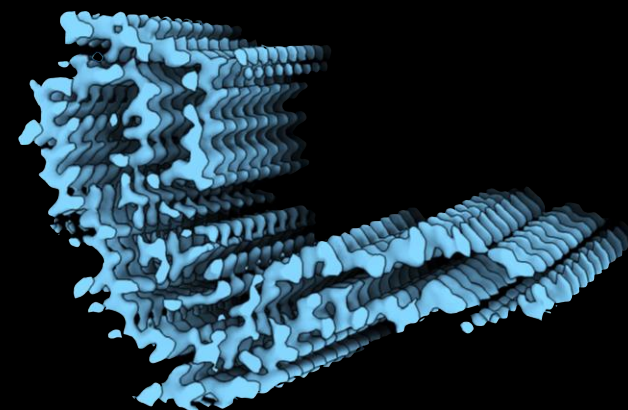
Manka et al. 2022
Hoyt et al. 2022

ME7



Manka et al. 2023

a22L



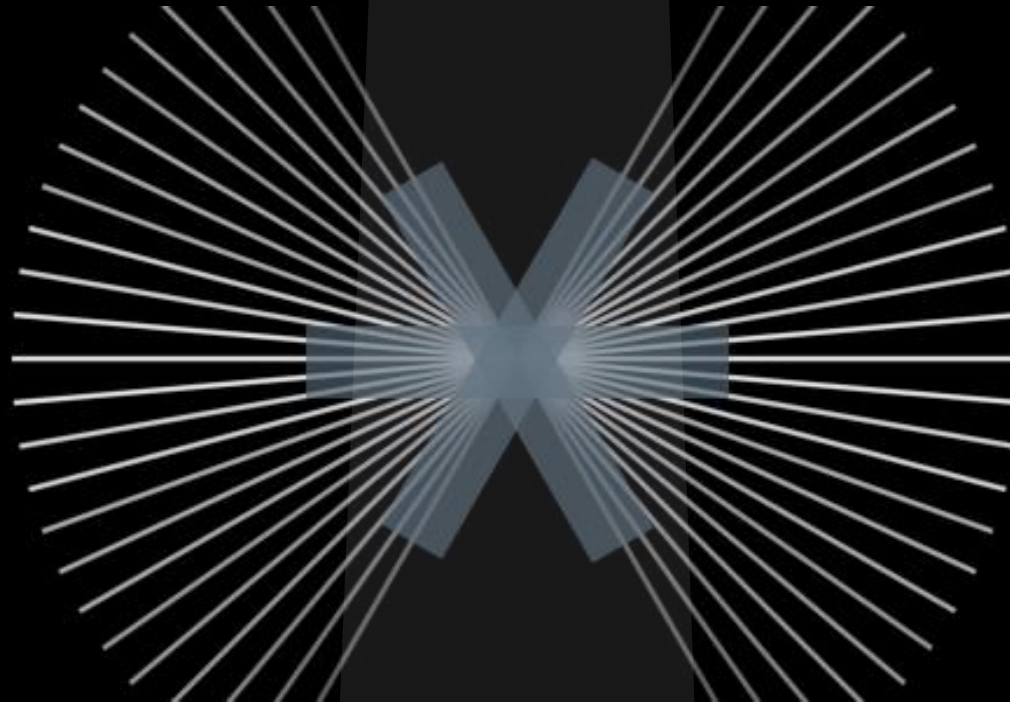
Hoyt et al. 2023

Cryo-electron tomography (Cryo-ET)

e- beam

60 °

2D sample
acquisition

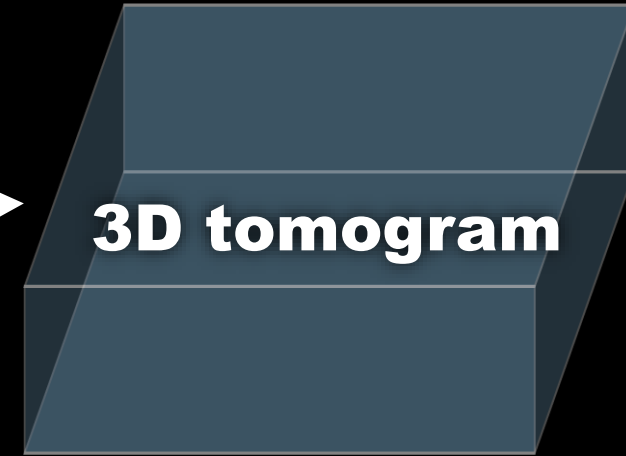


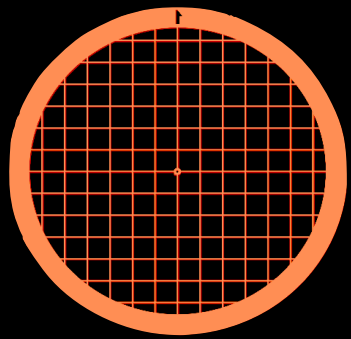
0 °



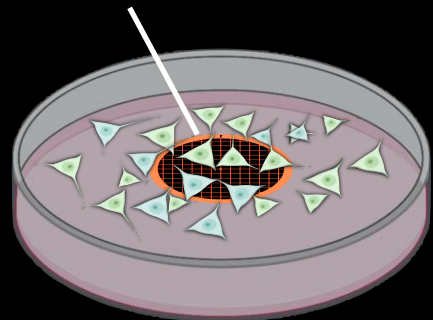
3D tomogram

-60 °

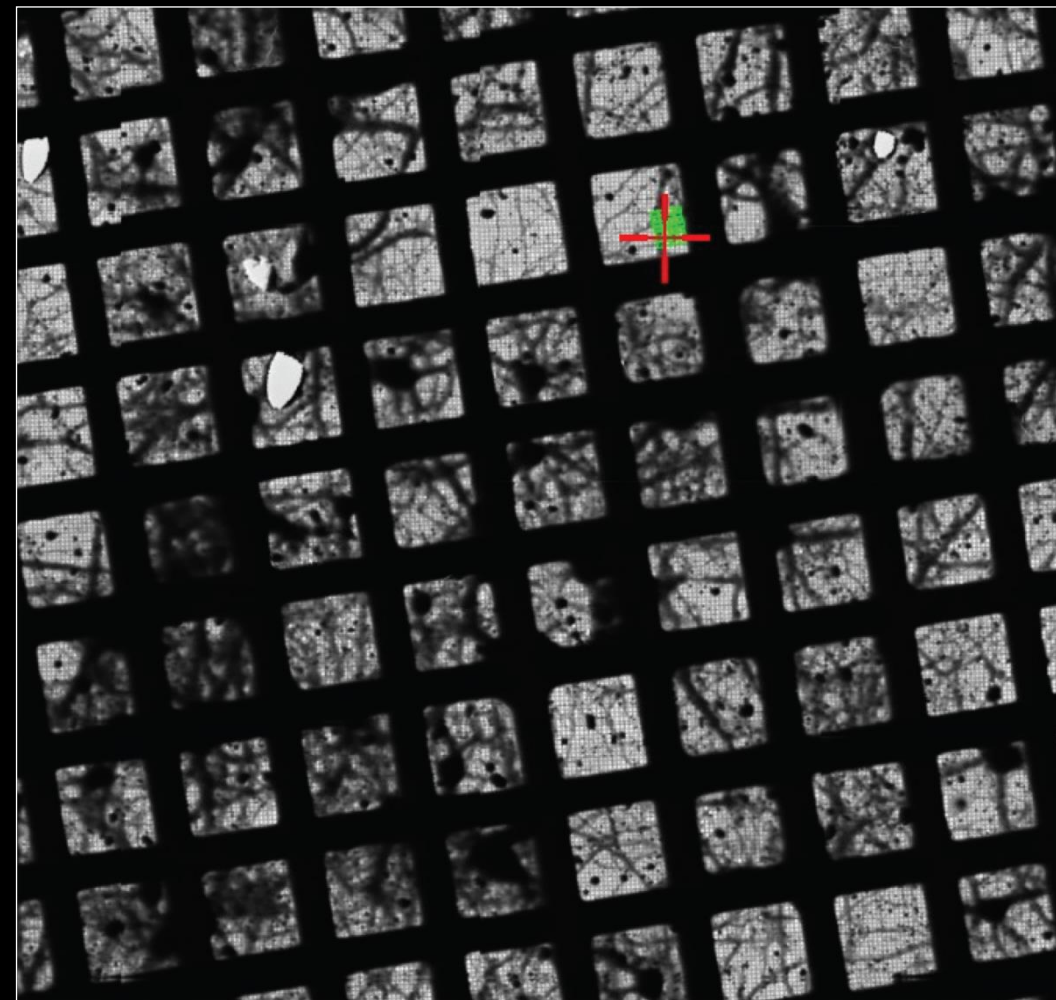




EM grid

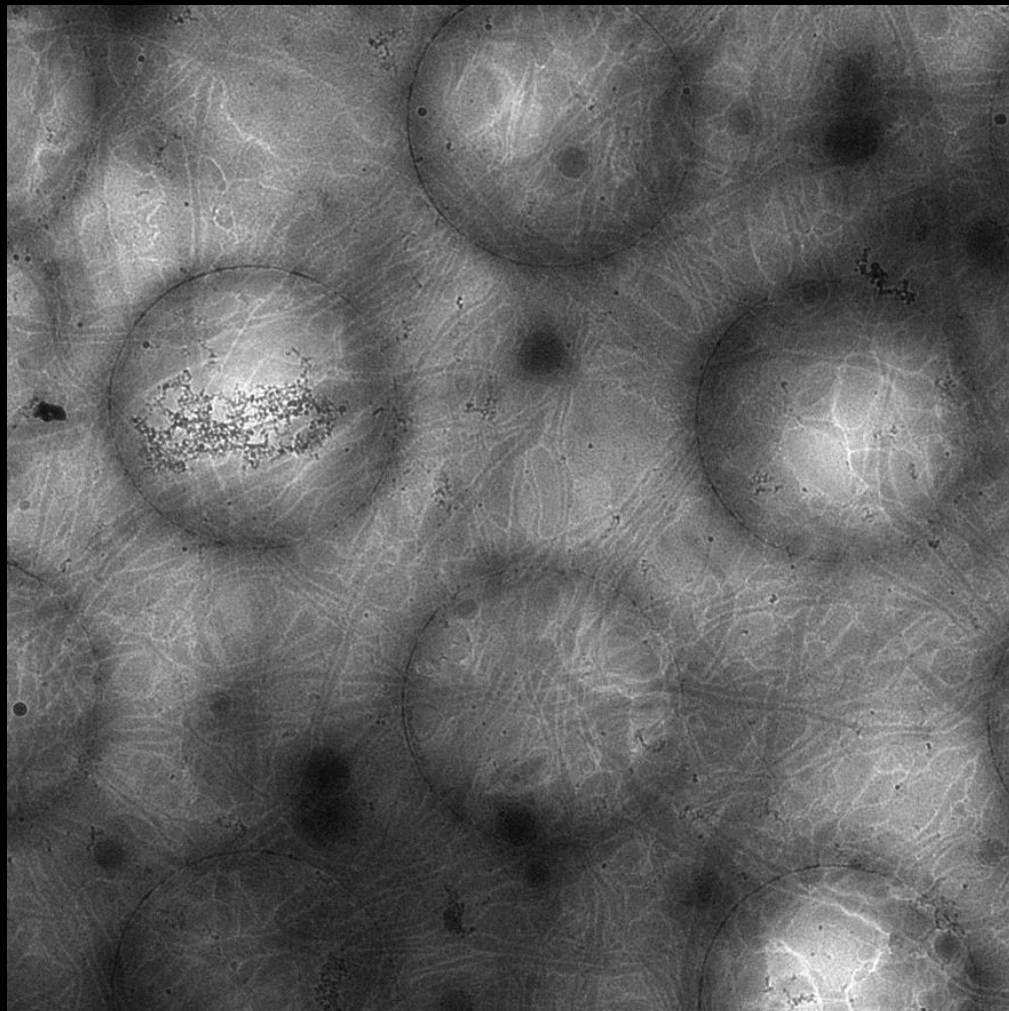


Primary neurons

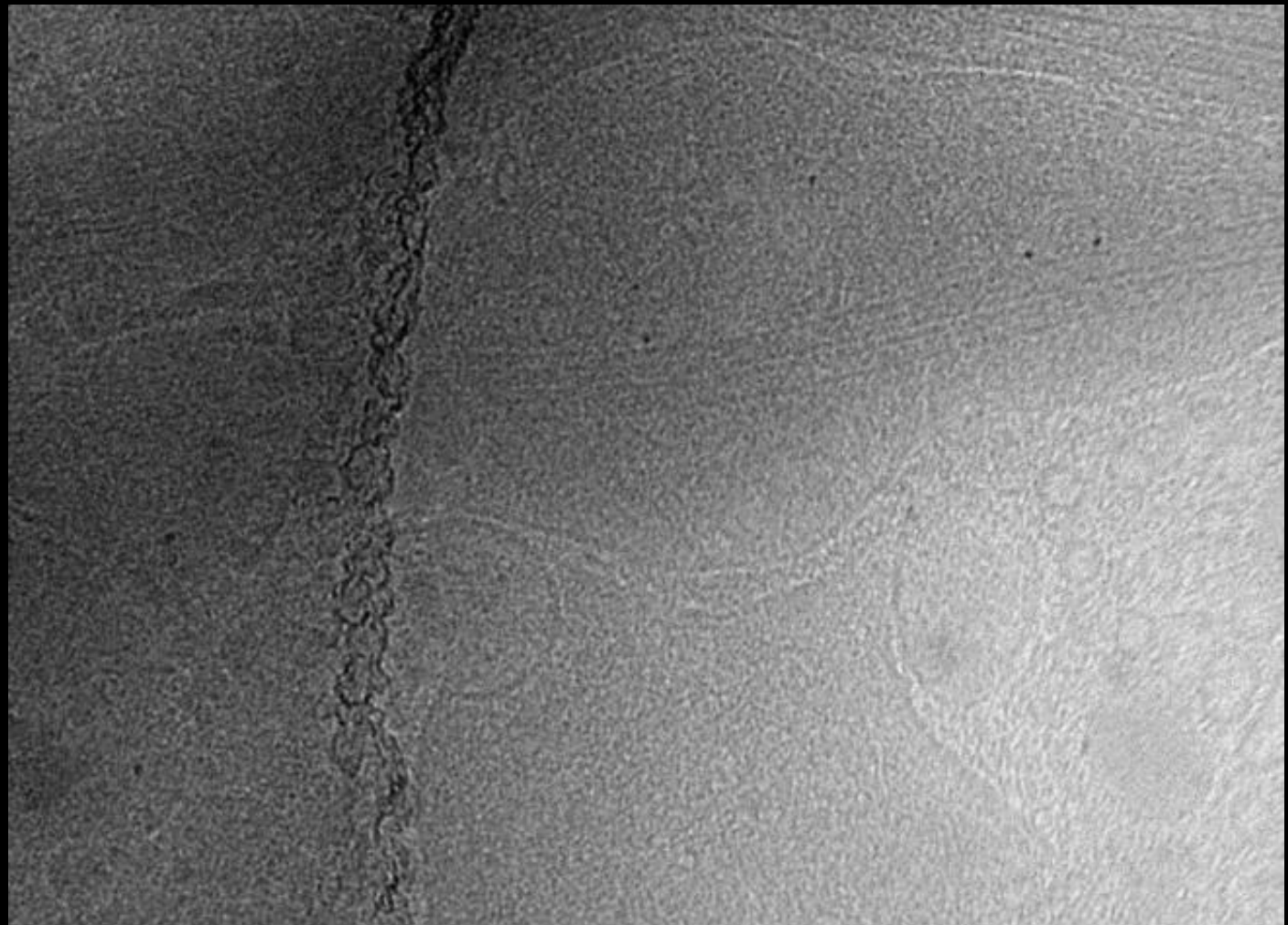


E17 mouse embryo





2 μm

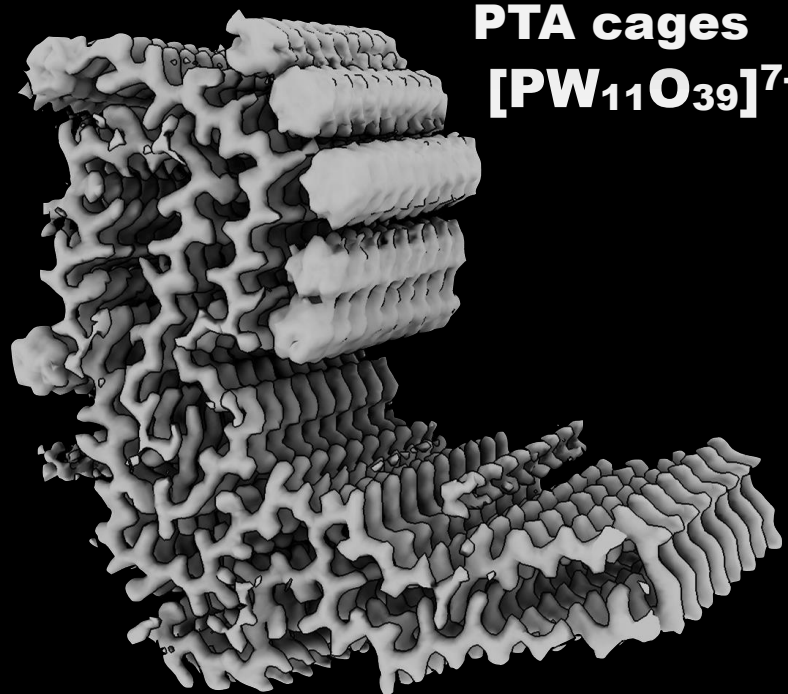


200 nm

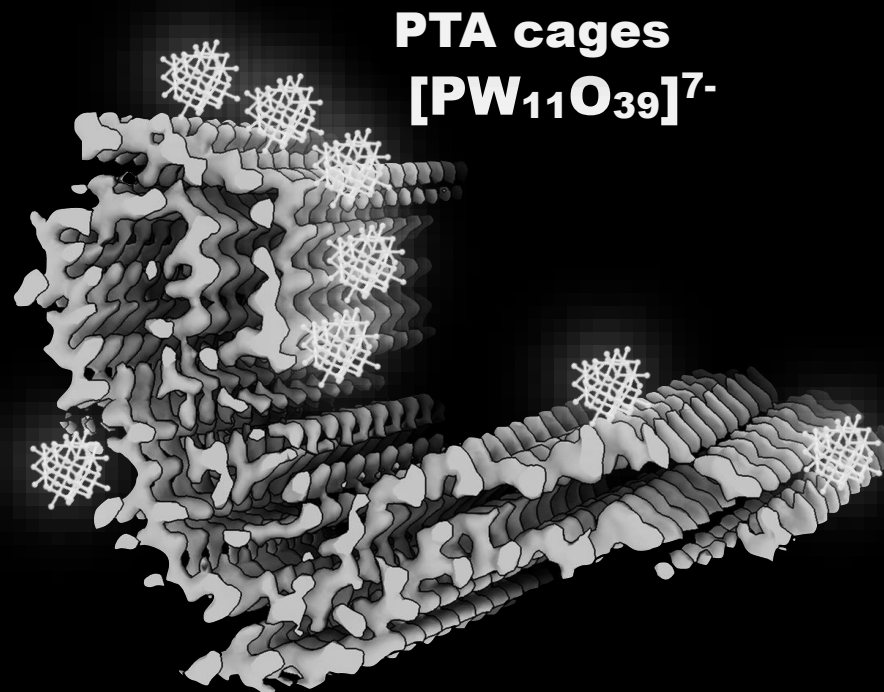
**Allows direct visualisation of
proteins within neurons**

But how to target prions?

RML



22L

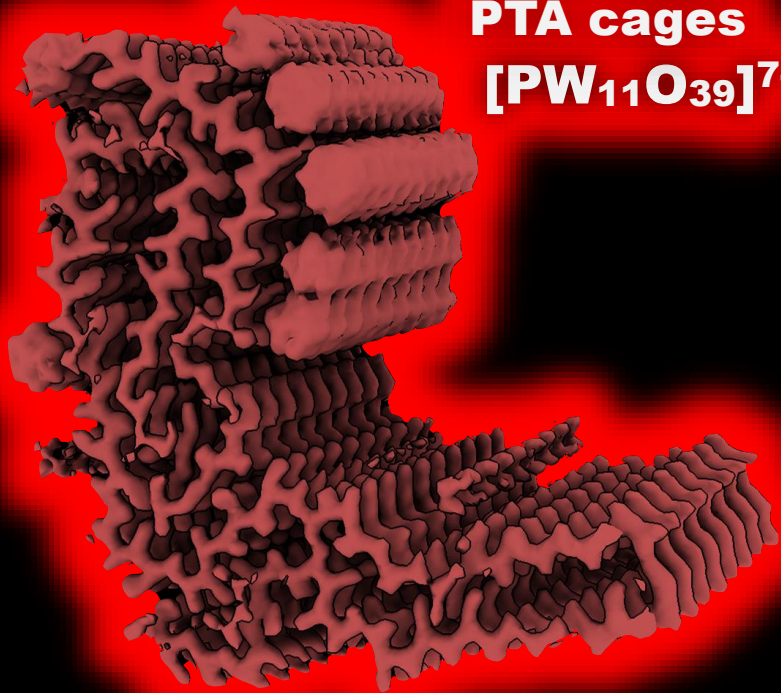


AlexaFluor546-NHS-ester

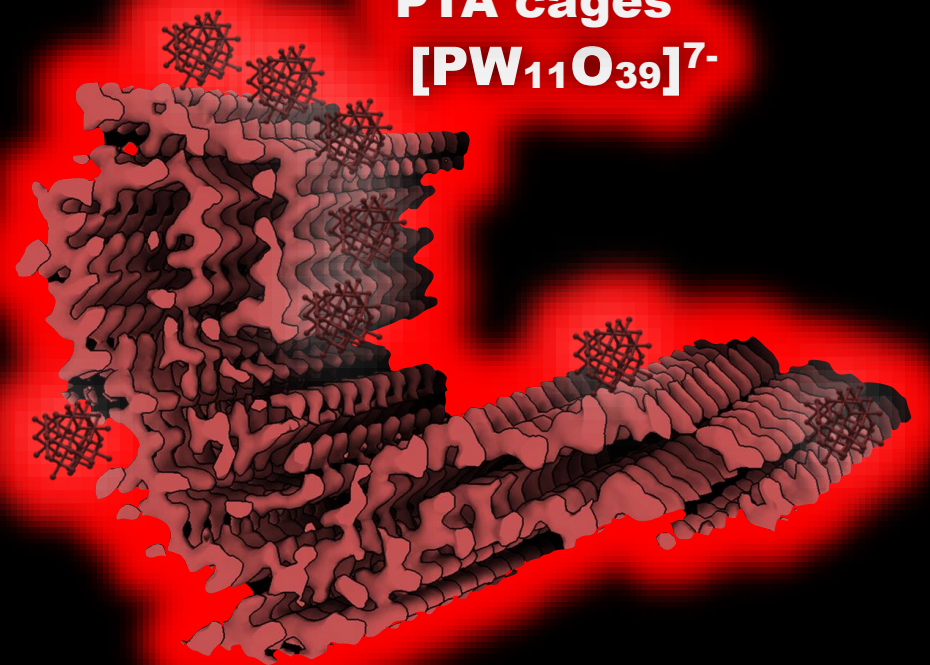
RML

+

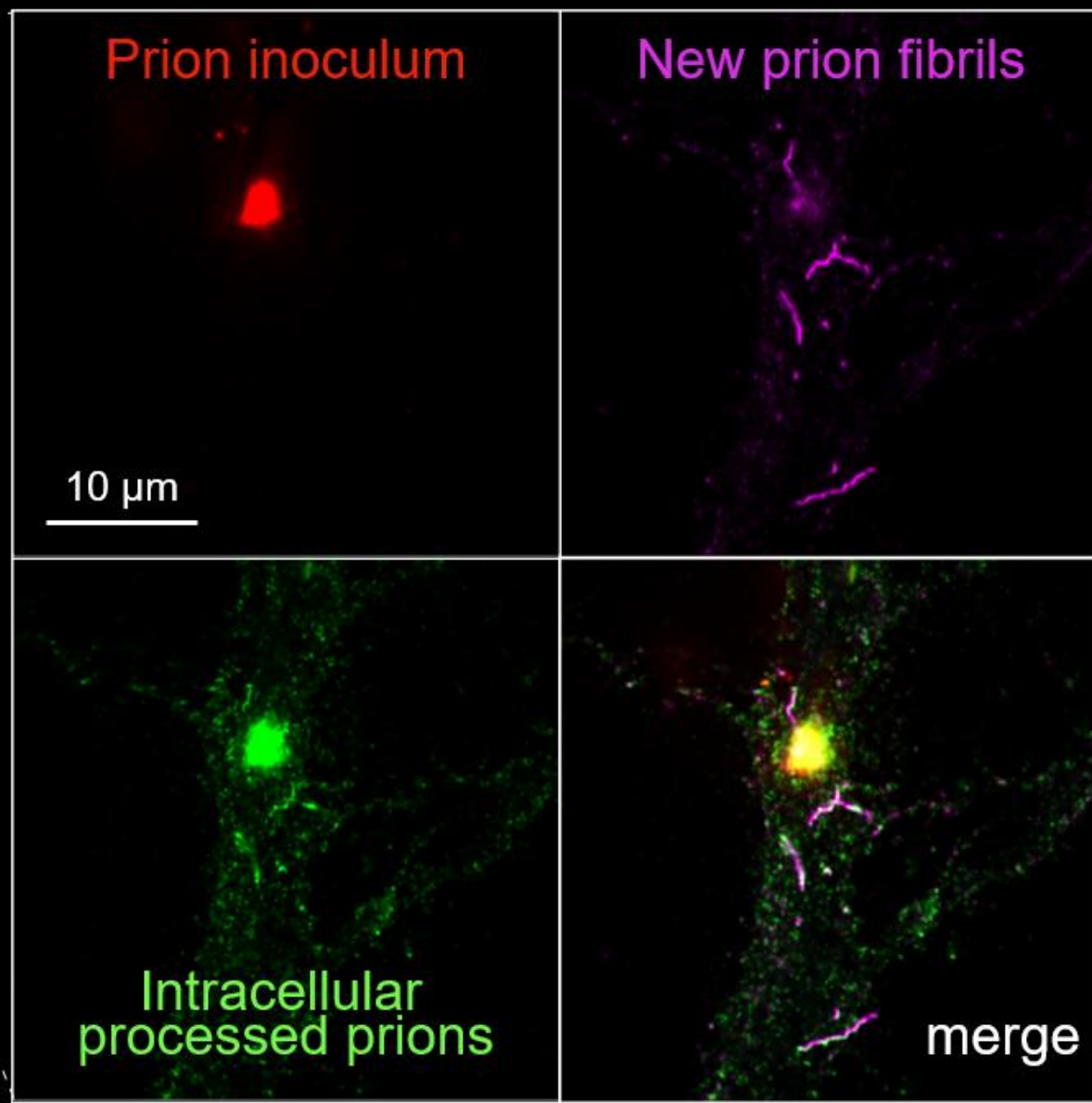
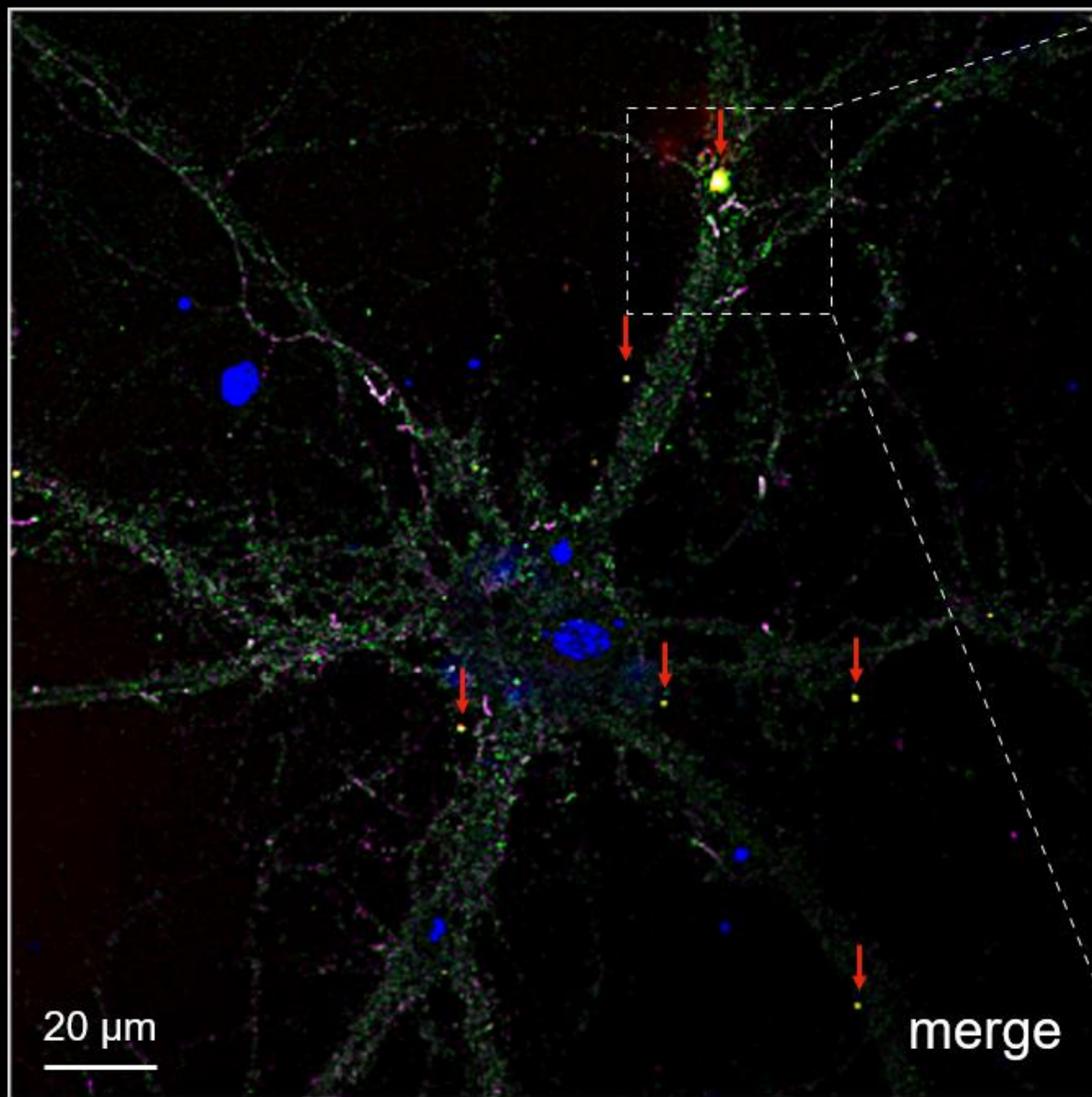
22L



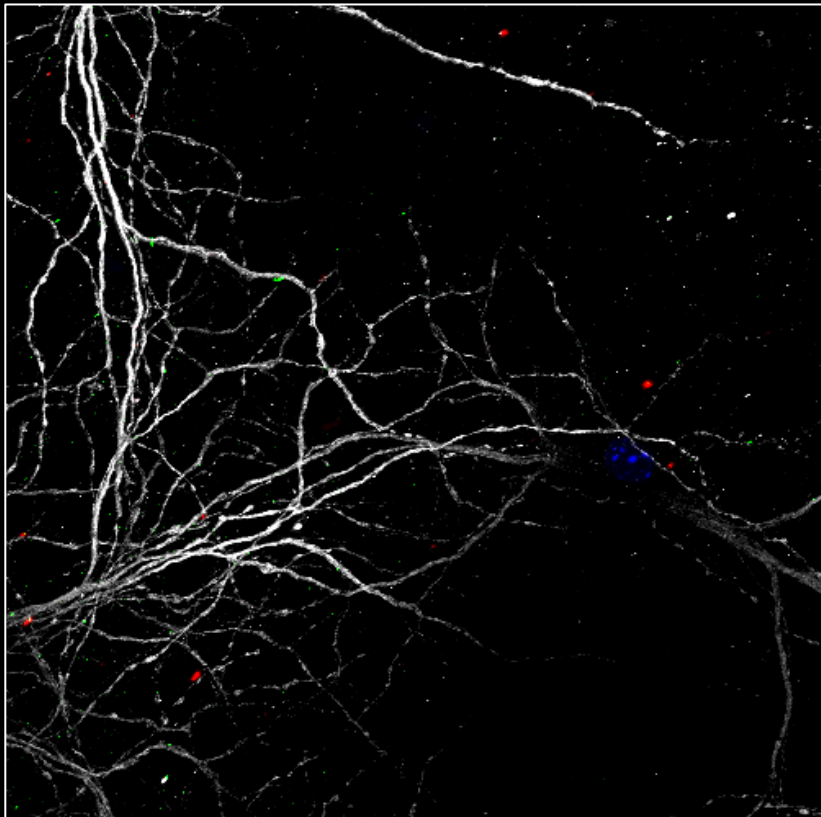
PTA cages
[PW₁₁O₃₉]⁷⁻



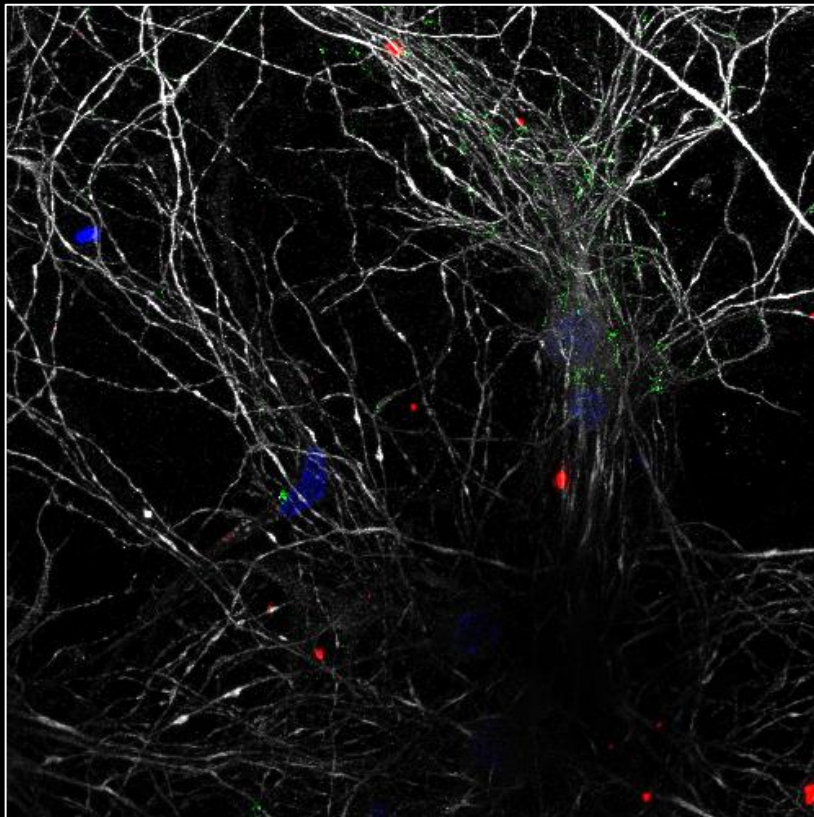
PTA cages
[PW₁₁O₃₉]⁷⁻



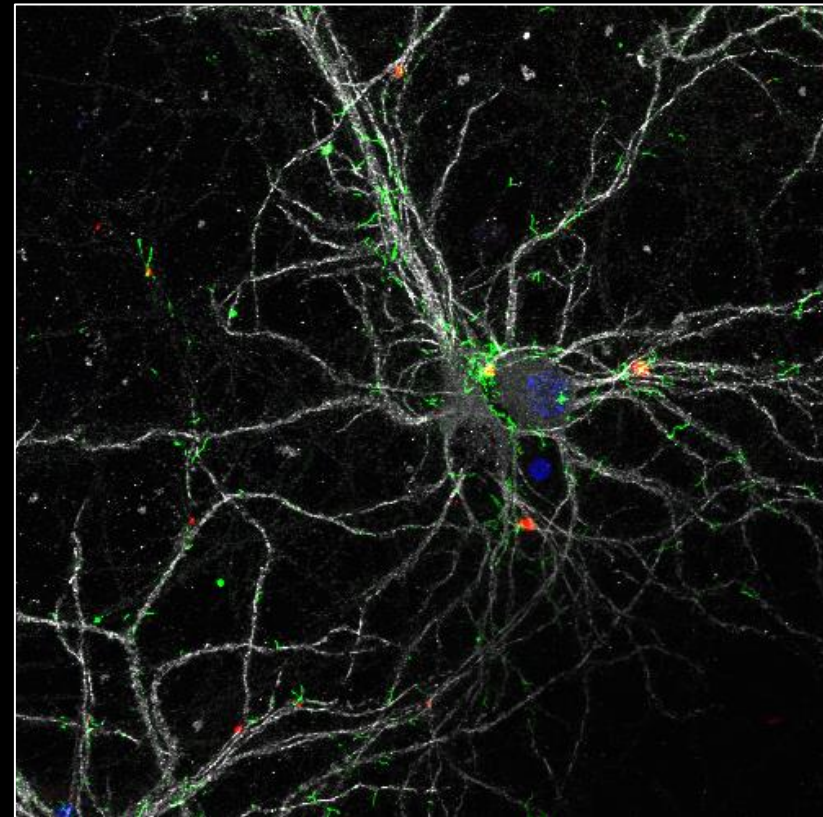
**7dpi
RML**



**14dpi
RML**



**21dpi
RML**



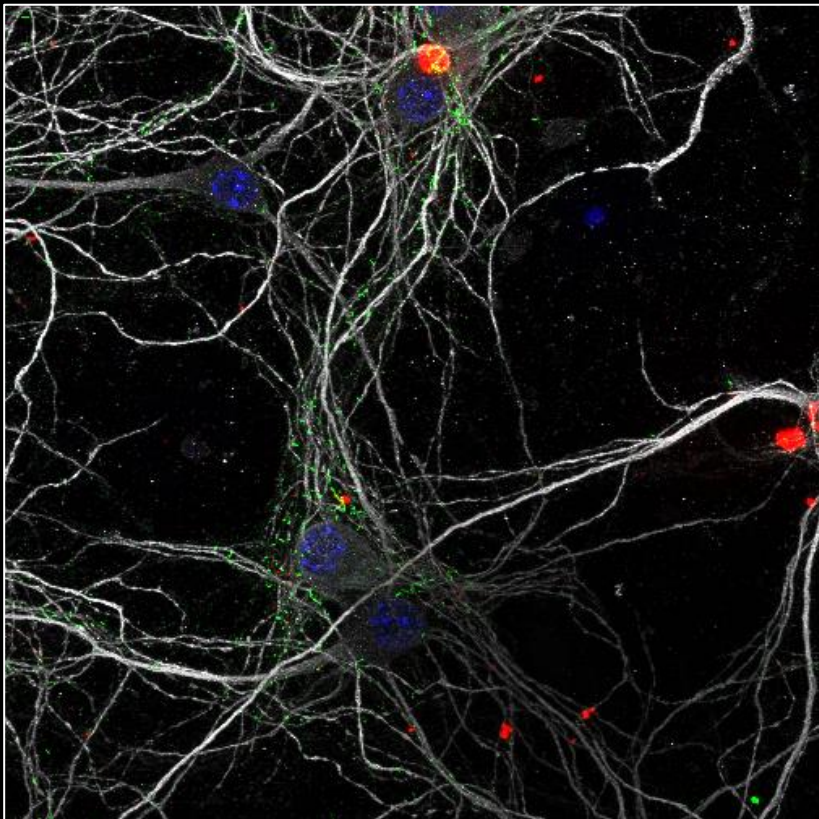
Fluorescence:

MAP2

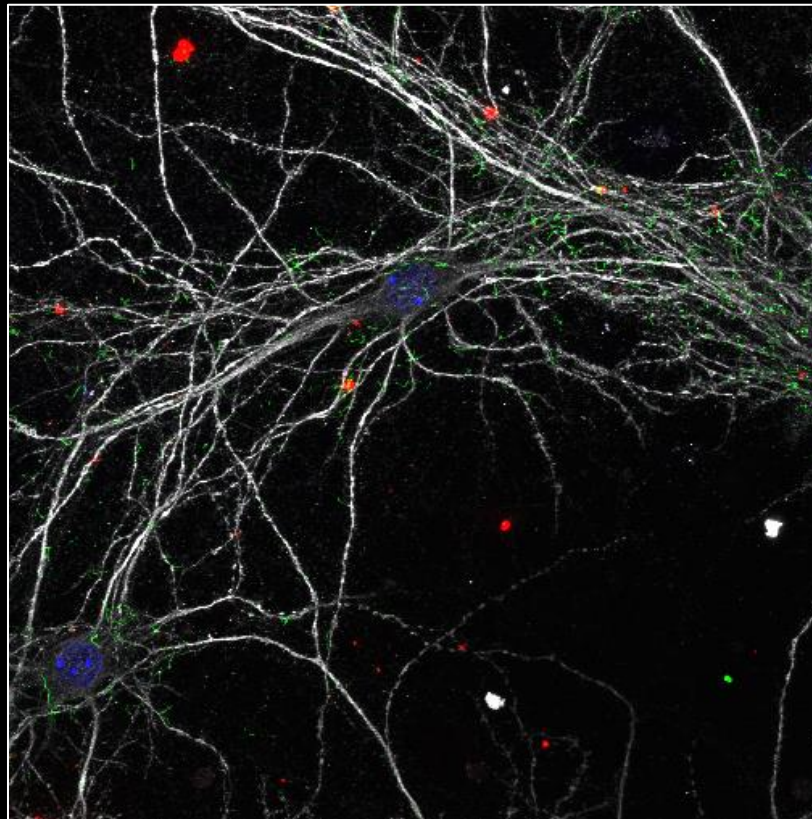
5B2

546-pRML

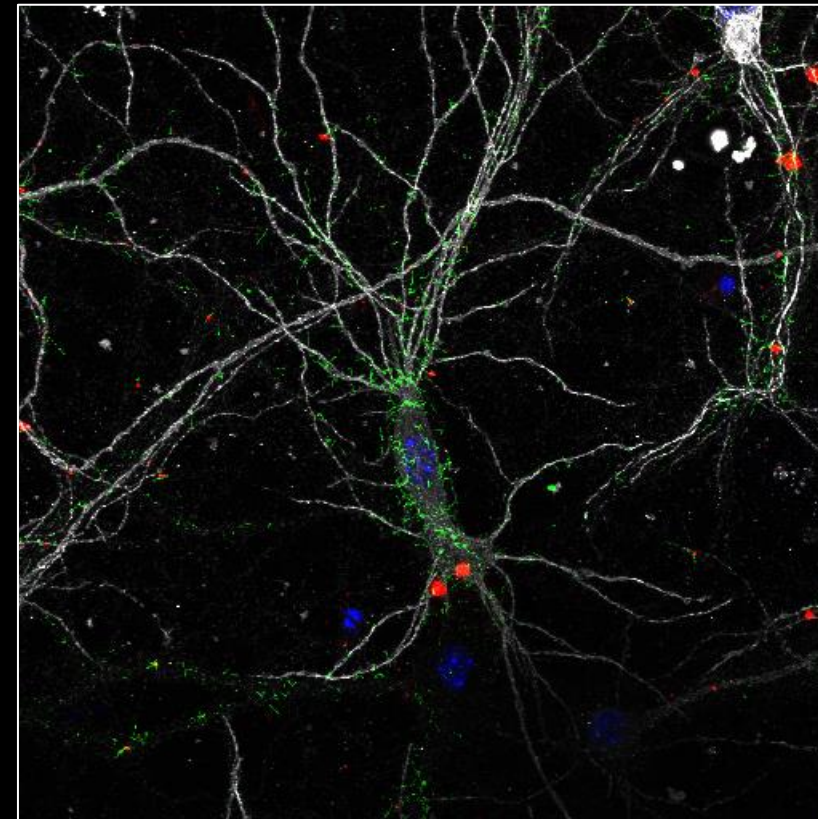
7dpi
22L



14dpi
22L



21dpi
22L



Fluorescence:

MAP2

5B2

546-pRML

Fluorescence:

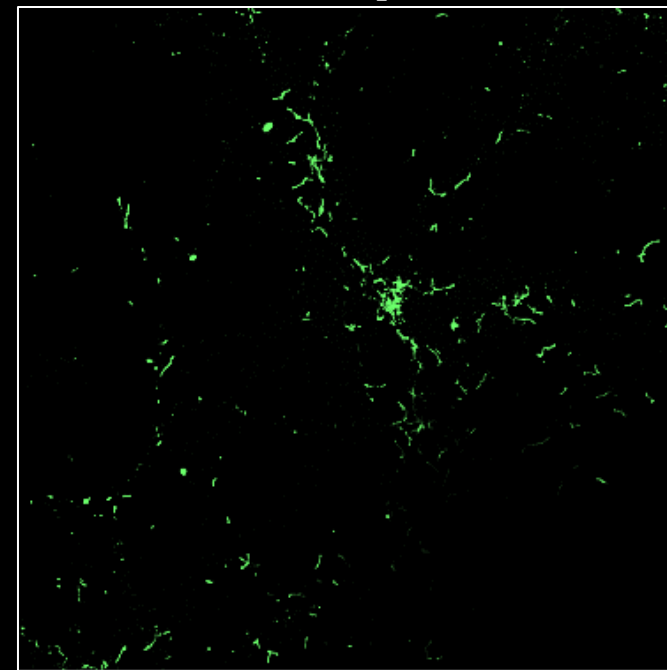
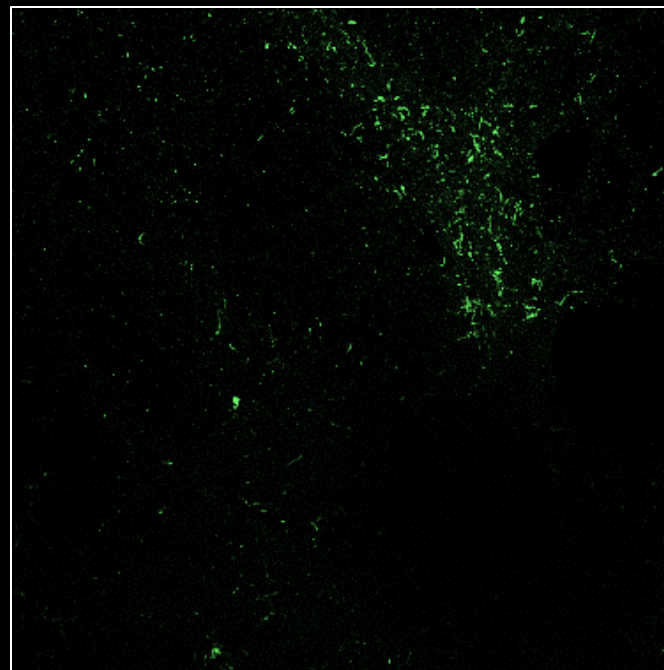
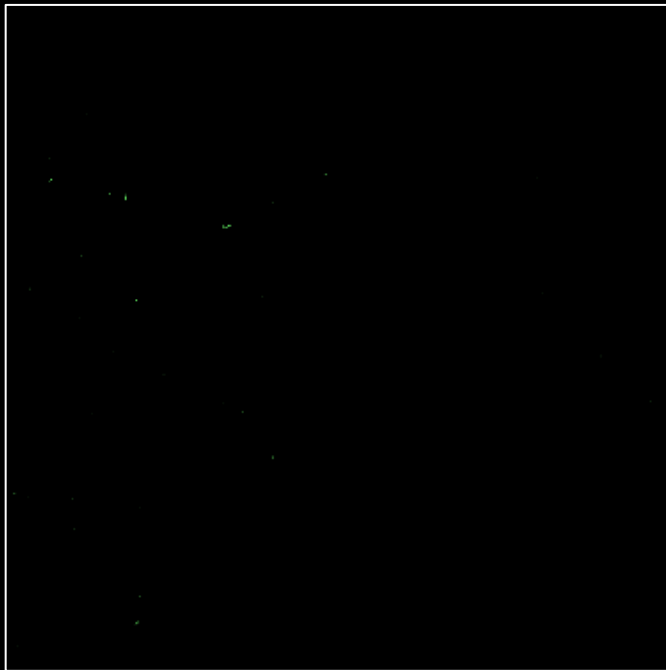
5B2

7dpi

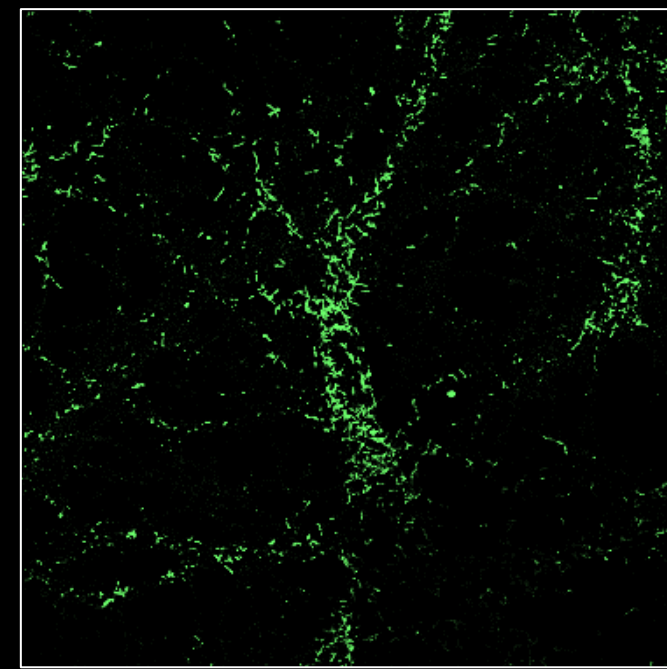
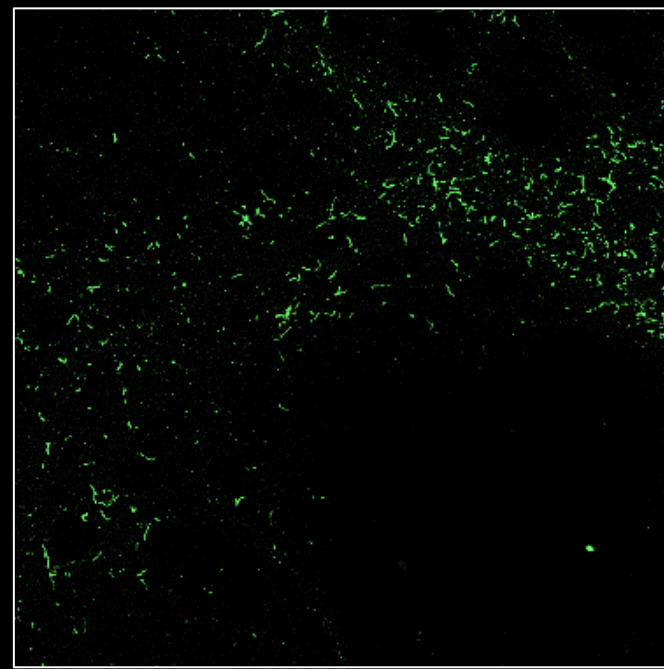
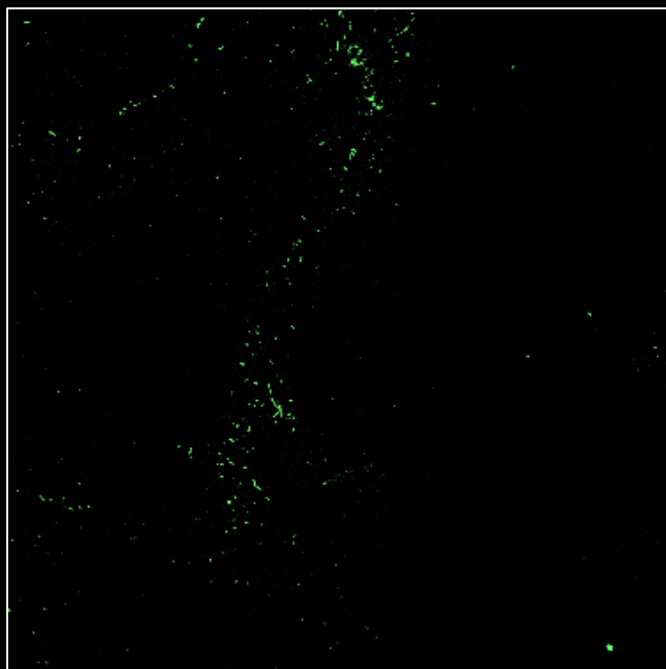
14dpi

21dpi

RML

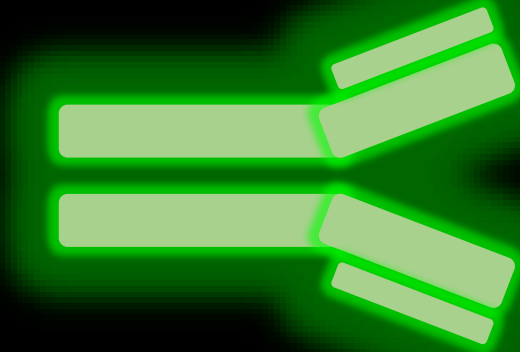


22L



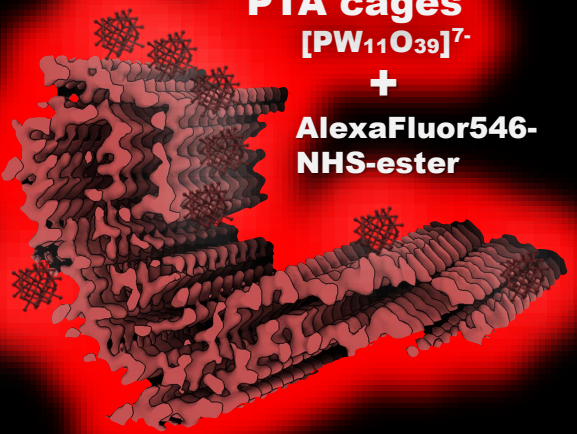
5B2

AlexaFluor488
primary
conjugation

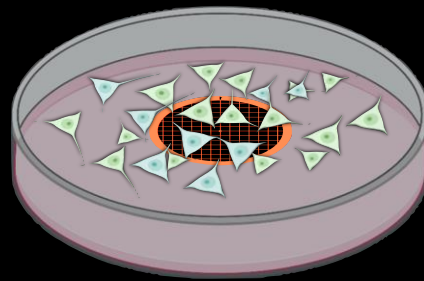


22L

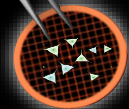
PTA cages
[PW₁₁O₃₉]⁷⁻
+
AlexaFluor546-
NHS-ester

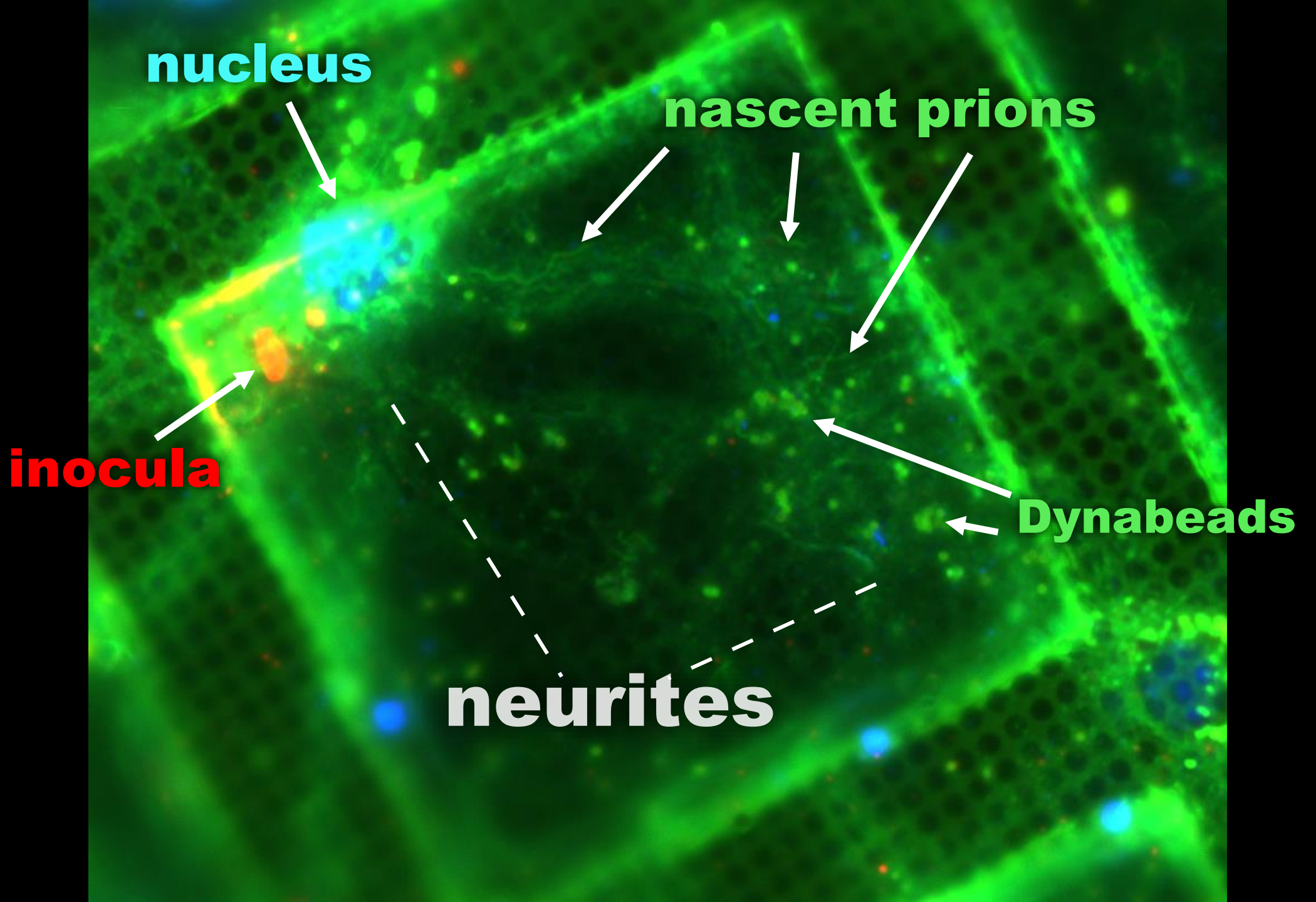


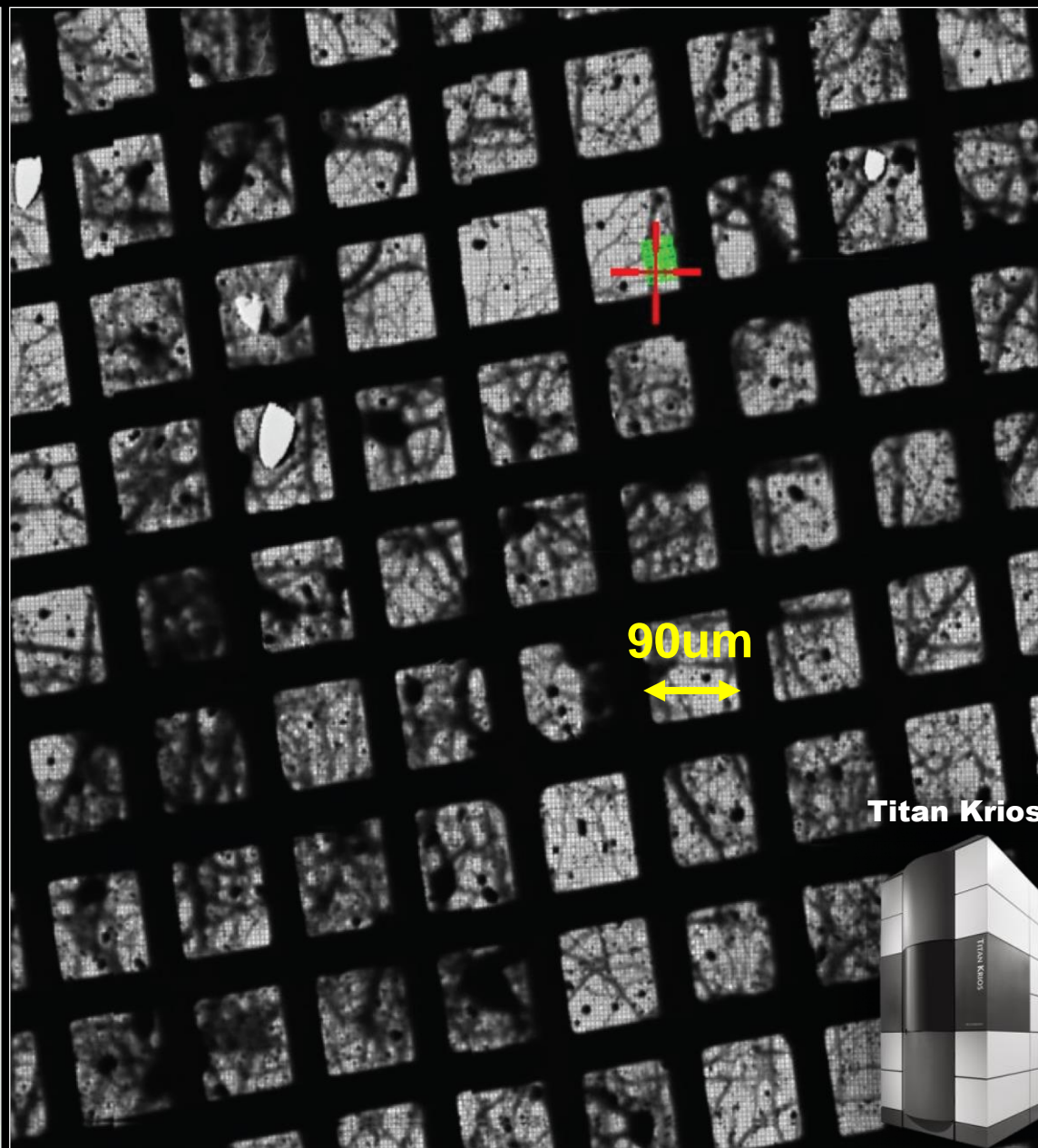
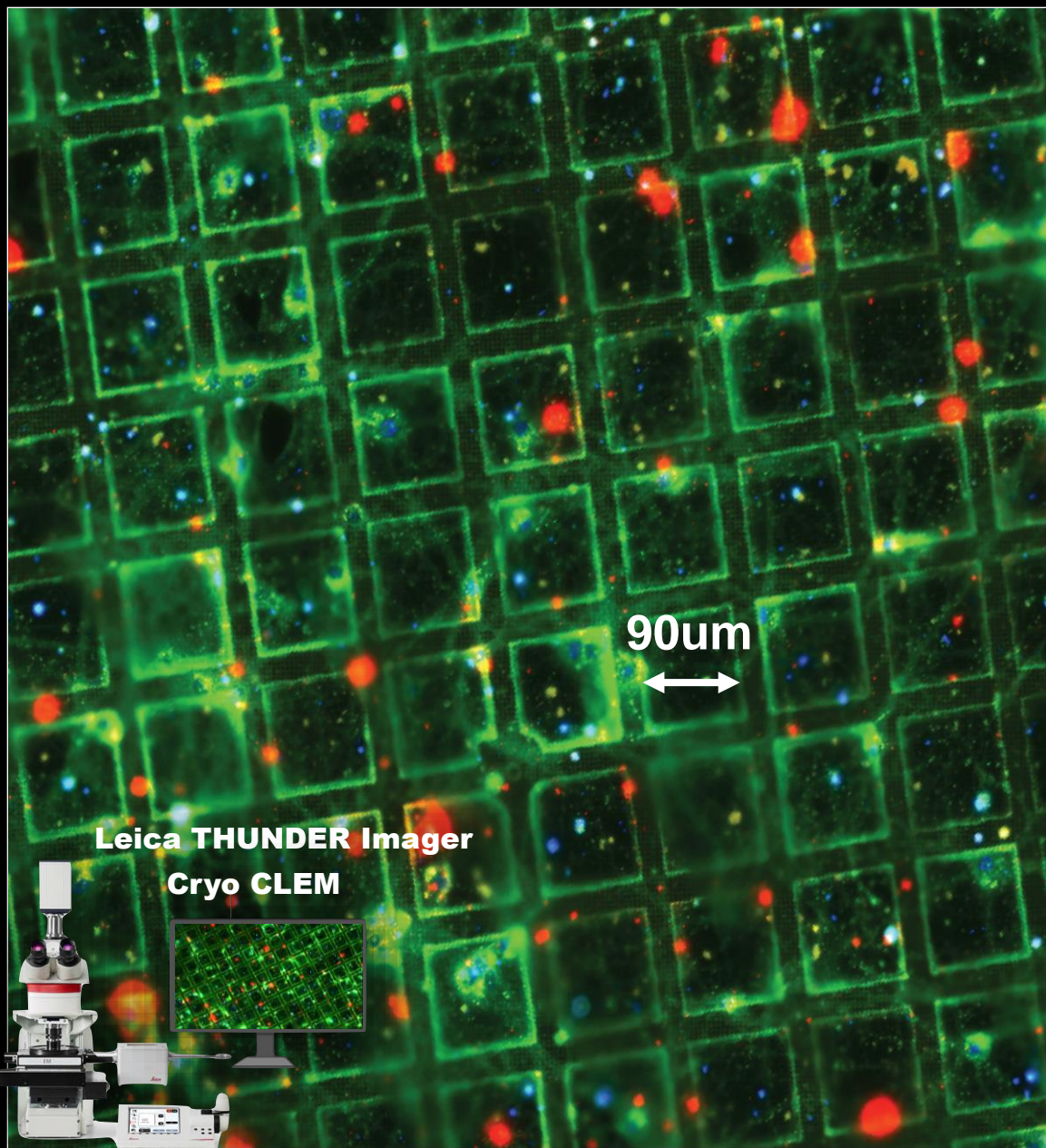
21dpi



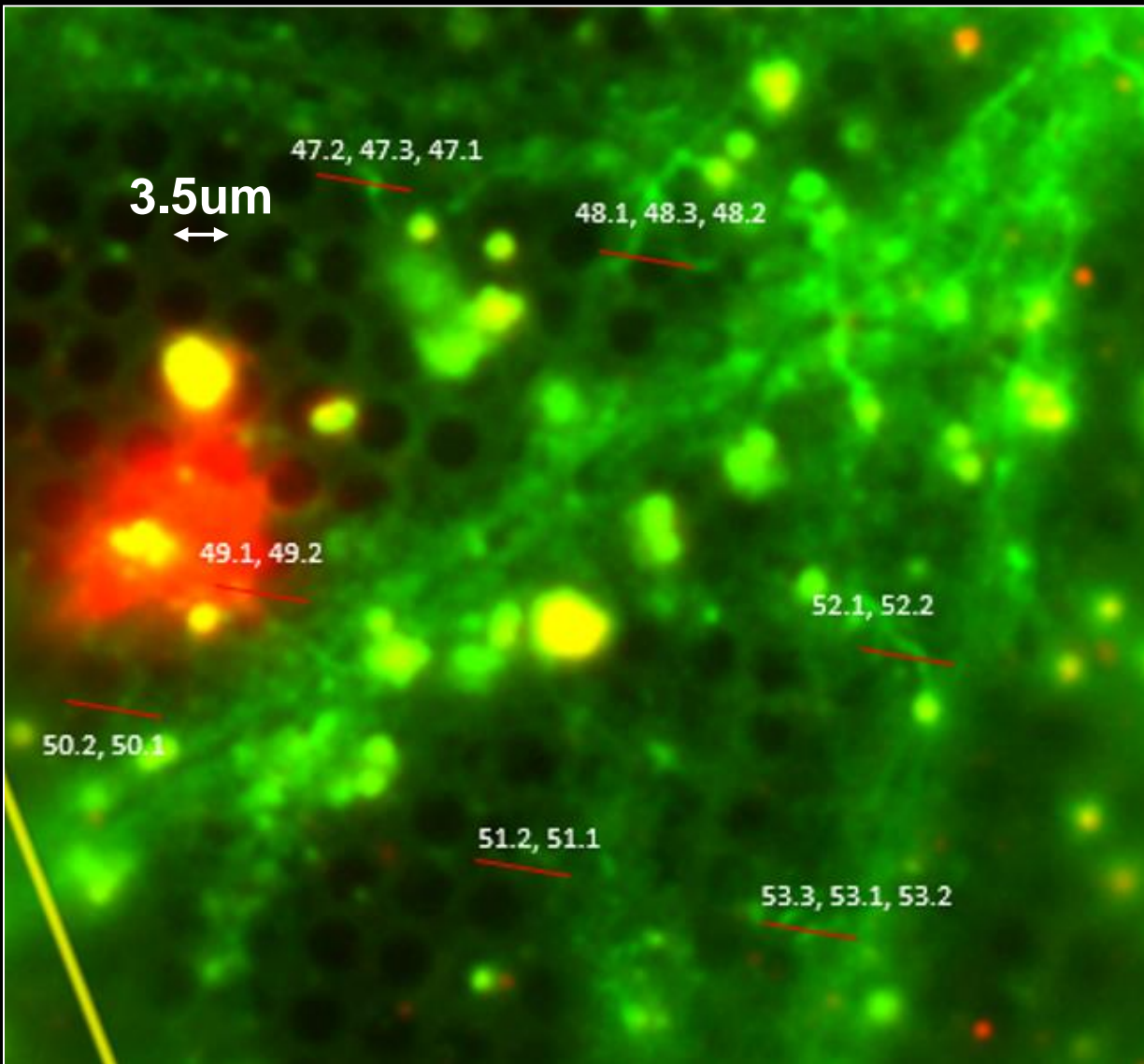
Plunge-freezing



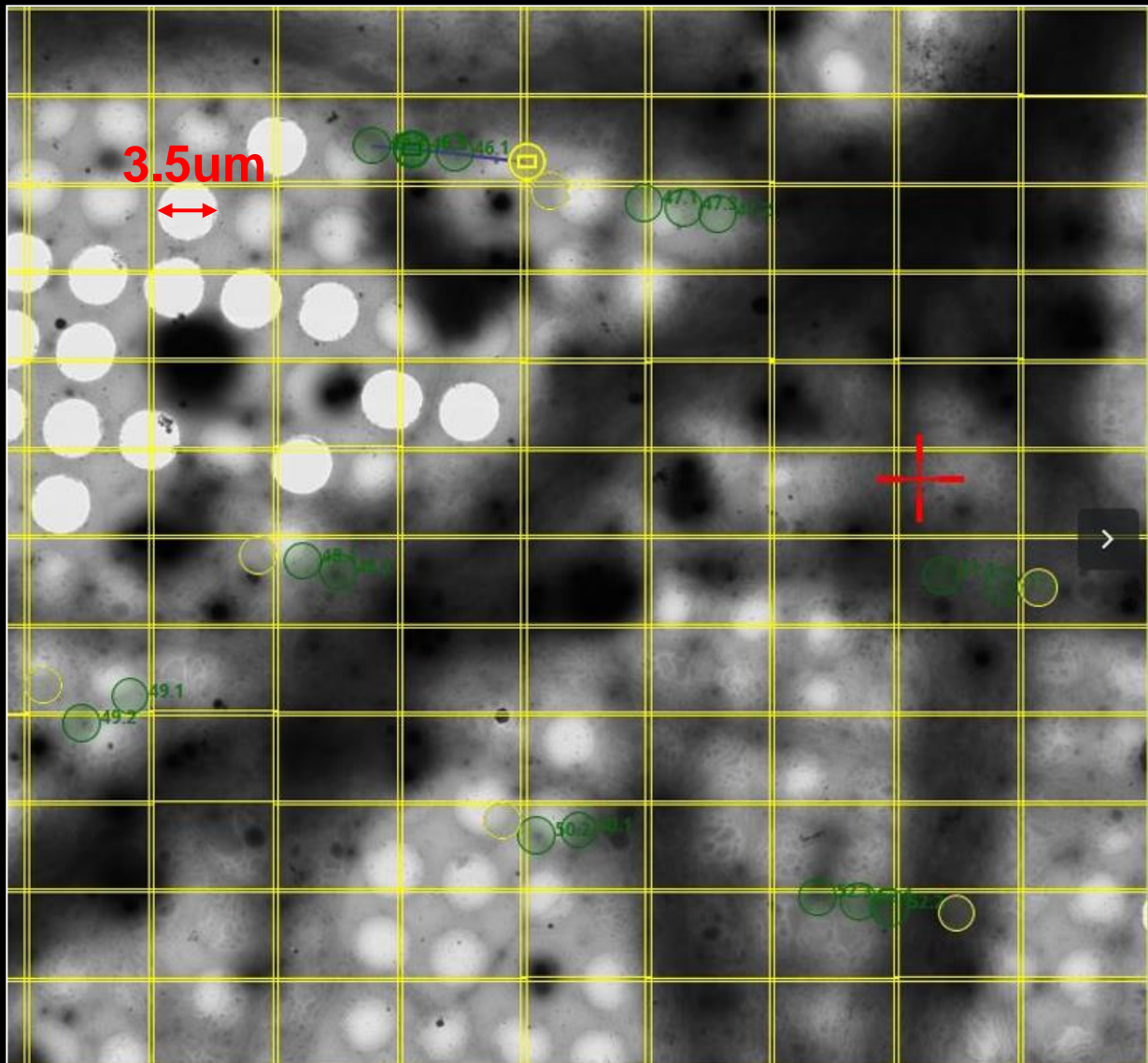




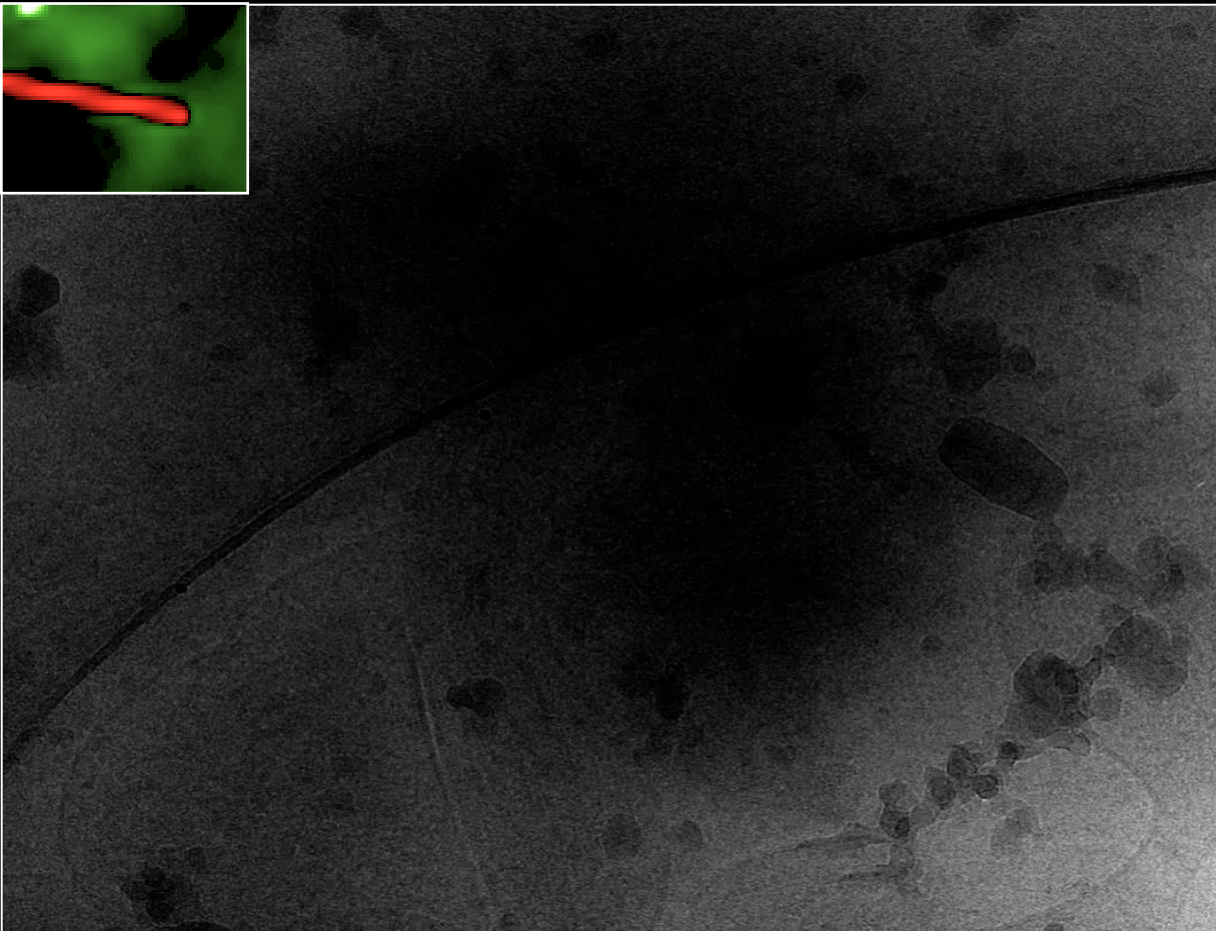
Cryo-widefield



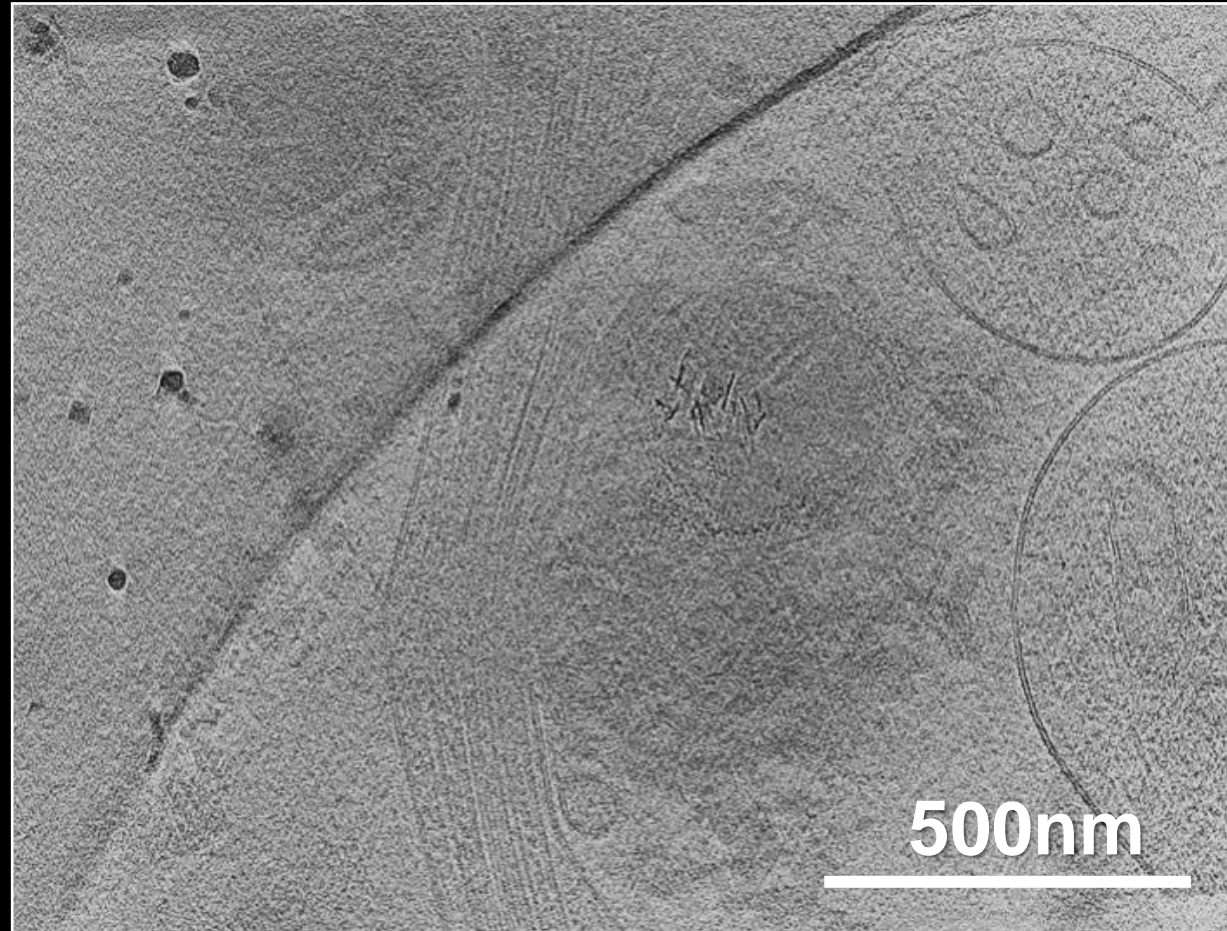
Cryo-EM

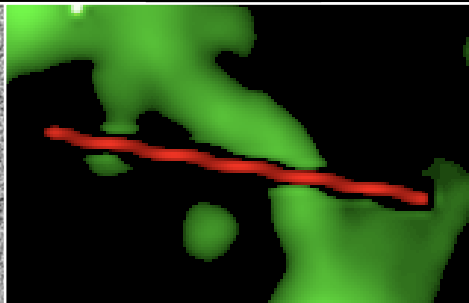
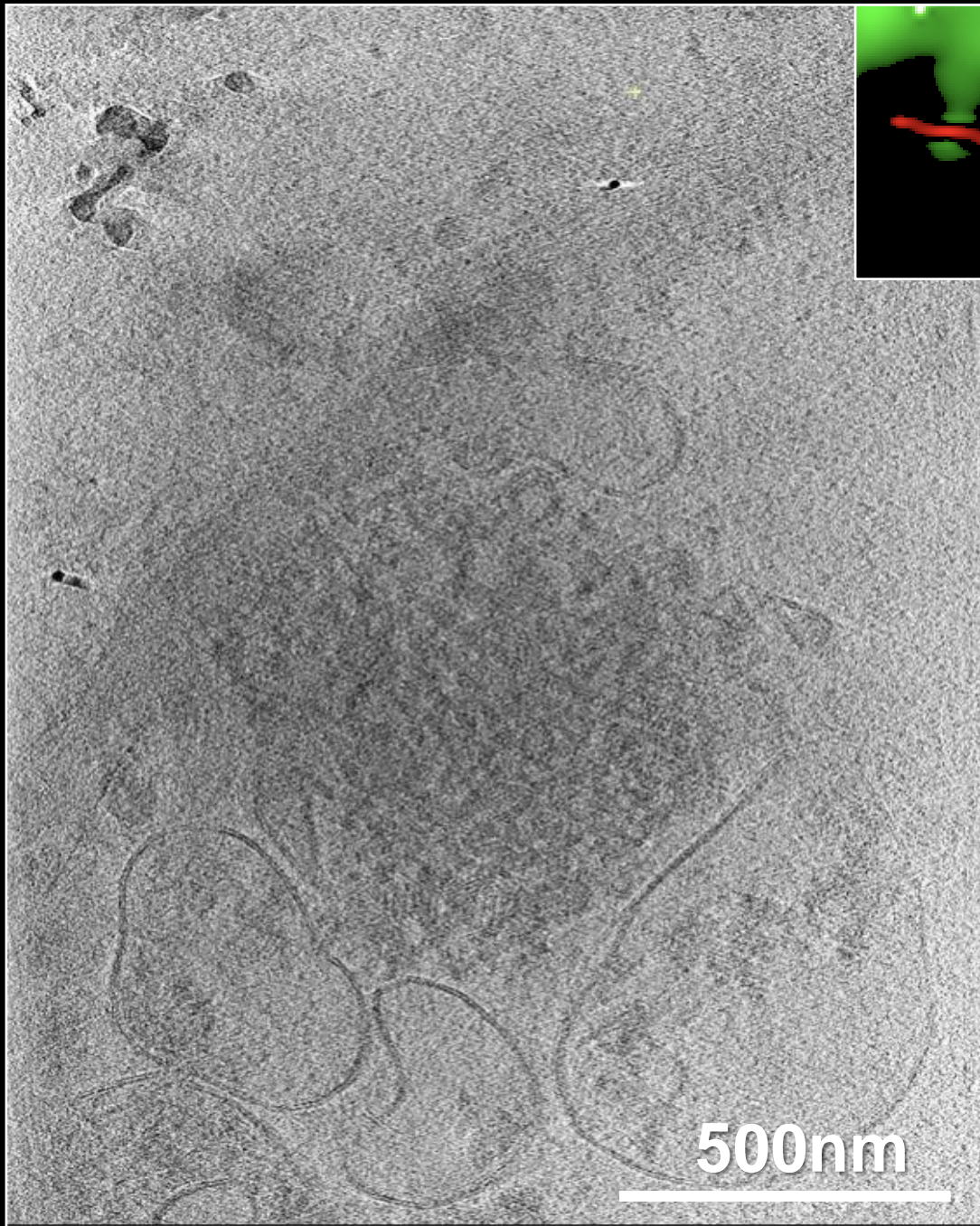


Tilt-series alignment



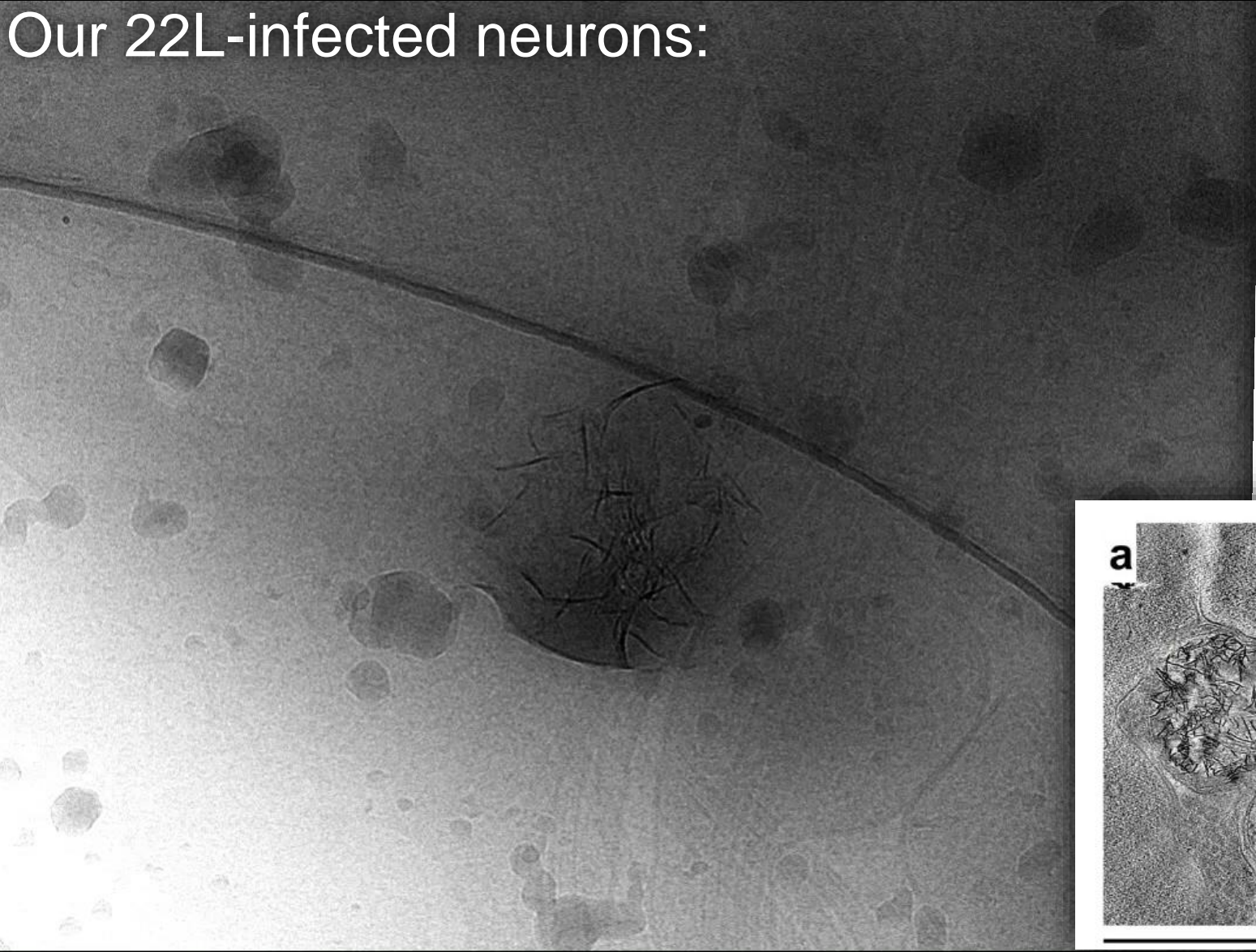
Reconstructed tomogram





Double membrane-bound compartments with sheet aggregates, shared in common with Huntington mutant and prion-infected neurons

Our 22L-infected neurons:



CryoET reveals organelle phenotypes in huntington disease patient iPSC-derived and mouse primary neurons

Received: 26 March 2022

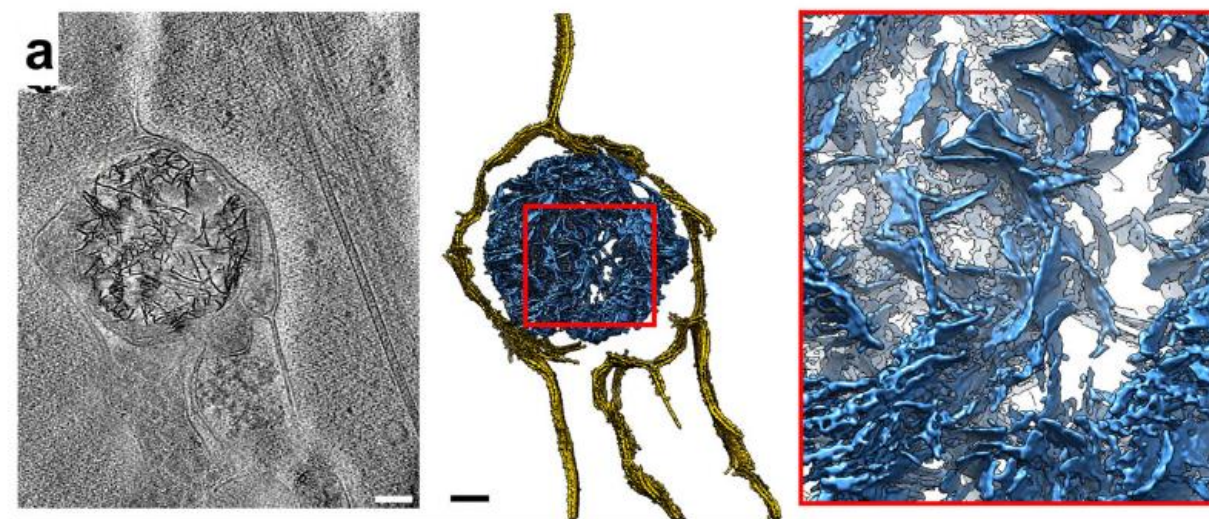
Accepted: 13 January 2023

Published online: 08 February 2023

Check for updates

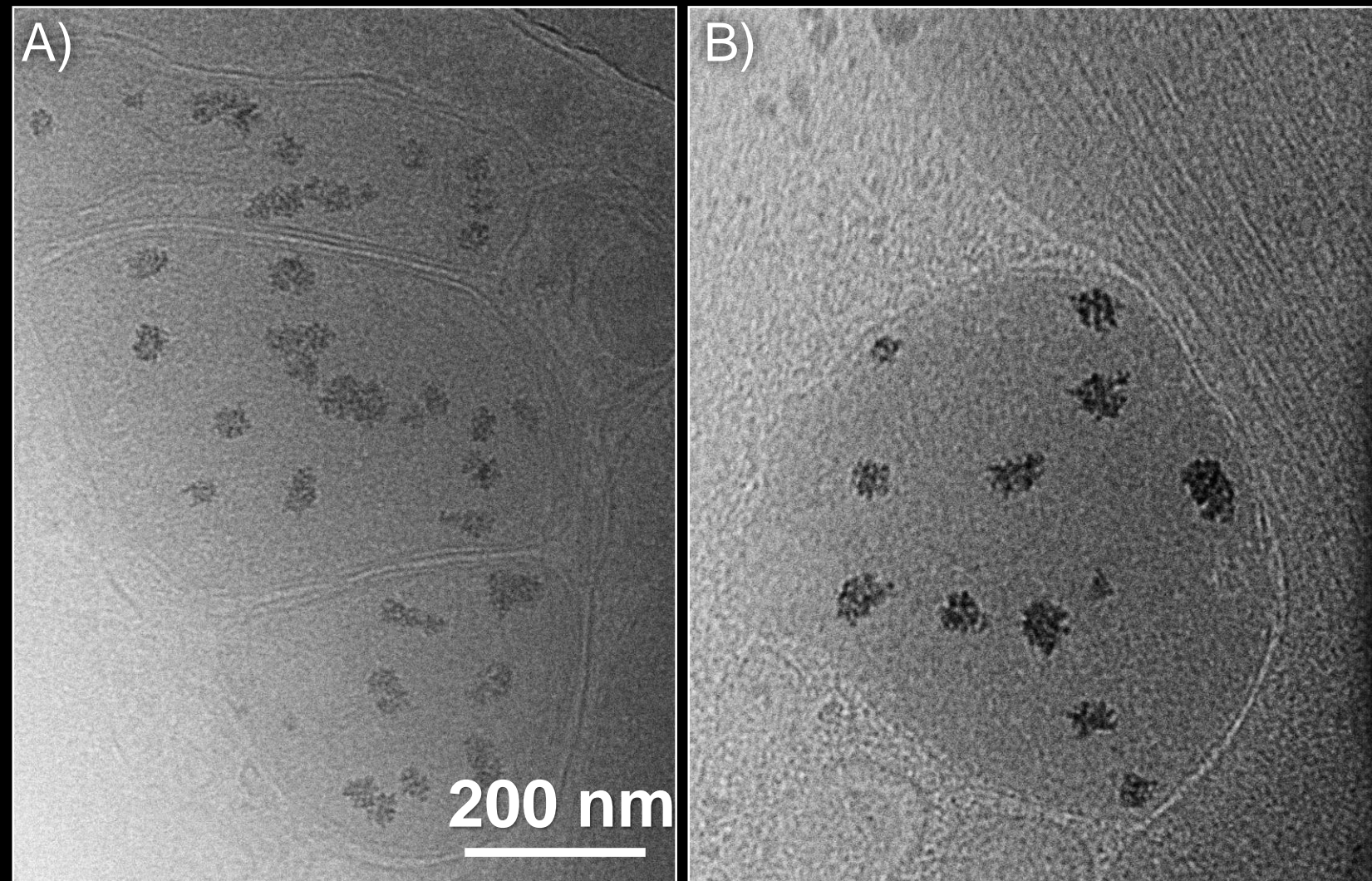
Gong-Her Wu^{1,14}, Charlene Smith-Geater^{2,14}, Jesús G. Galaz-Montoya¹, Yingli Gu³, Sanket R. Gupte⁴, Ranen Aviner⁵, Patrick G. Mitchell⁶, Joy Hsu⁴, Ricardo Miramontes⁷, Keona Q. Wang⁸, Nicolette R. Geller⁸, Cathy Hou¹, Cristina Danita¹, Lydia-Marie Joubert⁶, Michael F. Schmid⁶, Serena Yeung^{4,9}, Judith Frydman^{5,10}, William Mobley³, Chengbiao Wu³, Leslie M. Thompson^{2,7,8,11,12,15} & Wah Chiu^{1,6,13,15} ✉


Huntington's disease (HD) is caused by an expanded CAG repeat in the huntingtin gene, yielding a Huntingtin protein with an expanded polyglutamine tract. While experiments with patient-derived induced pluripotent stem cells (iPSCs) can help understand disease, defining pathological biomarkers remains challenging. Here, we used cryogenic electron tomography to visualize neurites in HD patient iPSC-derived neurons with varying CAG repeats, and primary cortical neurons from BACHD, deltaN17-BACHD, and wild-type mice. In HD models, we discovered sheet aggregates in double membrane-bound organelles, and mitochondria with distorted cristae and enlarged granules, likely mitochondrial RNA granules. We used artificial intelligence to quantify mitochondrial granules, and proteomics experiments reveal differential protein content in isolated HD mitochondria. Knockdown of Protein Inhibitor of Activated STAT1 ameliorated aberrant phenotypes in iPSC- and BACHD neu-



Calcium phosphate granules accumulate in mitochondria, and cristae become poorly defined

22L-infected neurons:



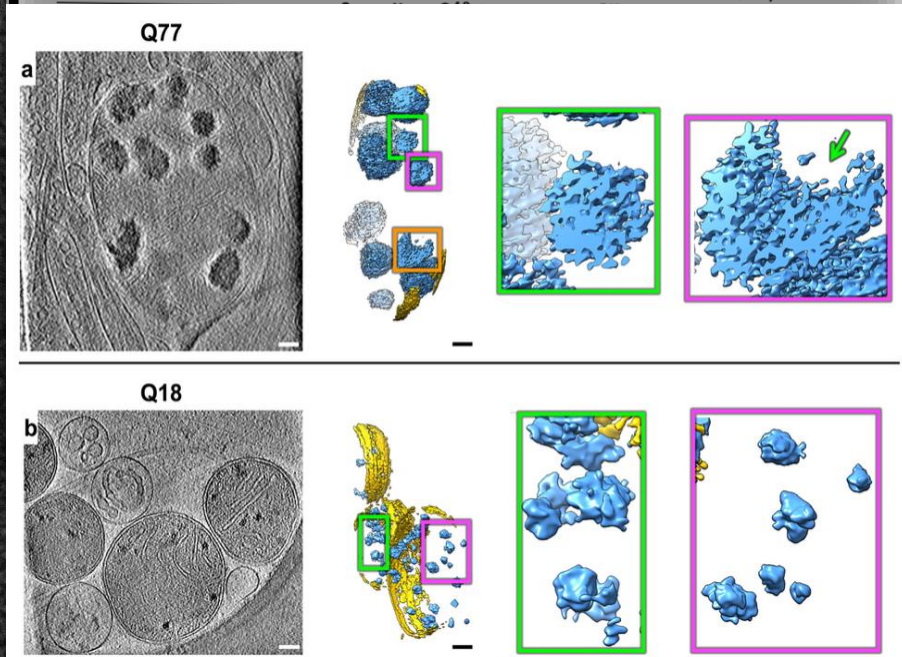
nature communications 

Article <https://doi.org/10.1038/s41467-023-36096-w>

CryoET reveals organelle phenotypes in huntington disease patient iPSC-derived and mouse primary neurons

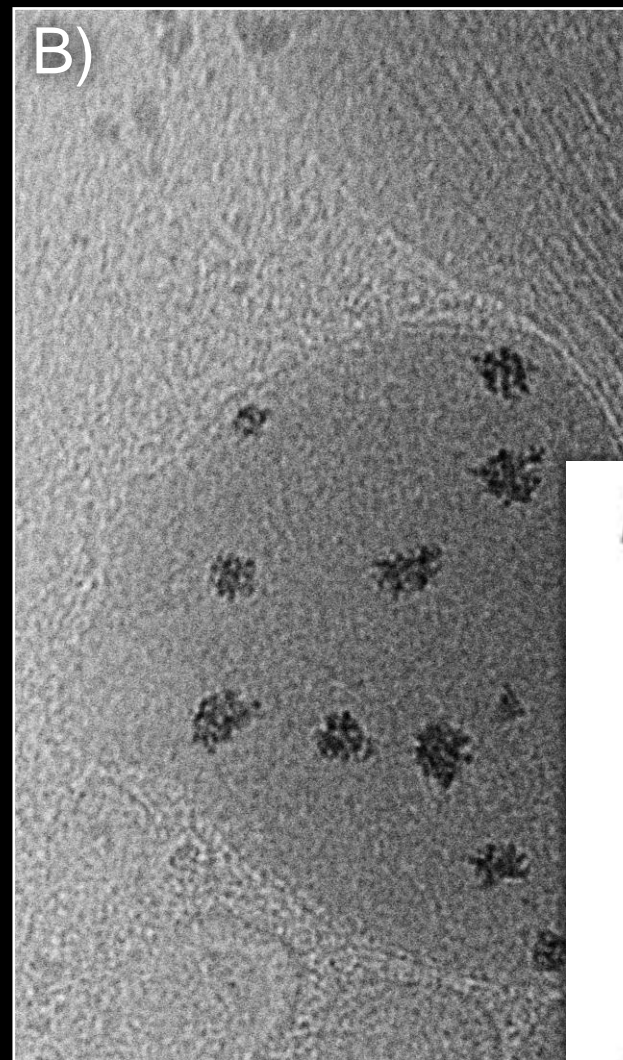
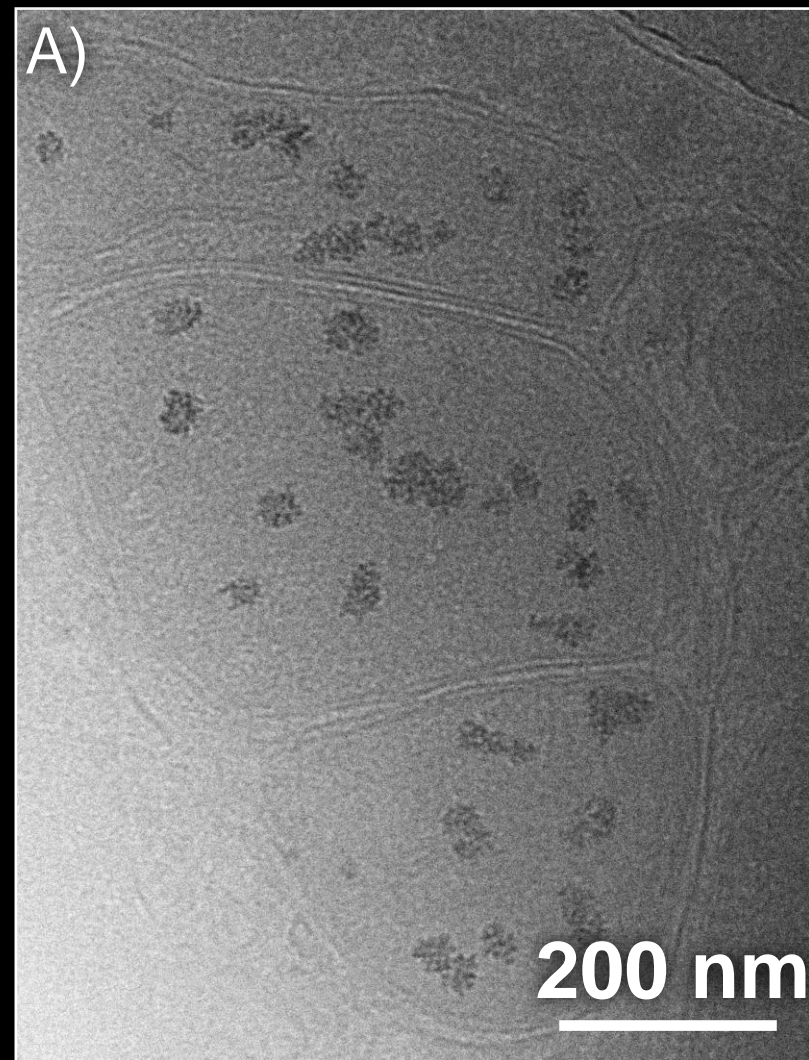
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Gong-Her Wu^{1,14}, Charlene Smith-Geater^{2,14}, Jesús G. Galaz-Montoya¹, Yingli Gu³, Sanket R. Gupte⁴, Ranen Aviner⁵, Patrick G. Mitchell⁶, Joy Hsu⁴, Ricardo Miramontes⁷, Keona Q. Wang⁸, Nicolette R. Geller⁹, Cathy Hou¹, Cristina Danita¹, Lydia-Marie Joubert¹⁰, Michael F. Schmid⁶



Calcium phosphate granules accumulate in mitochondria, and cristae become poorly defined

22L-infected neurons:



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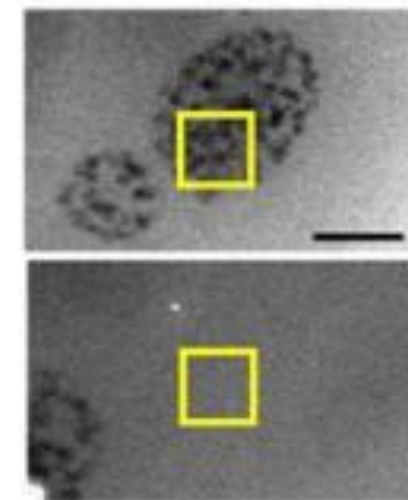
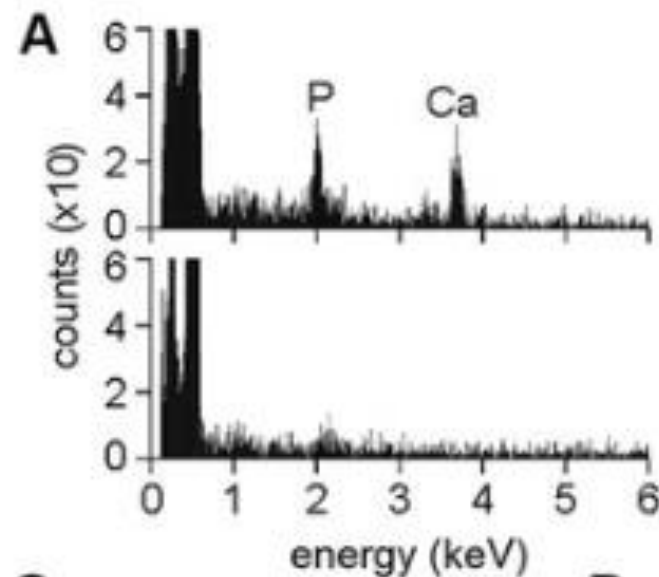
Research Article
Cell Biology

3D visualization of mitochondrial solid-phase calcium stores in whole cells

Sharon Grayer Wolf , Yael Mutsafi, Tali Dadosh, Tal Ilani, Zipora Lansky, Ben Horowitz, Sarah Rubin, Michael Elbaum, Deborah Fass 

Weizmann Institute of Science, Israel

Nov 6, 2017 • <https://doi.org/10.7554/eLife.29929>  

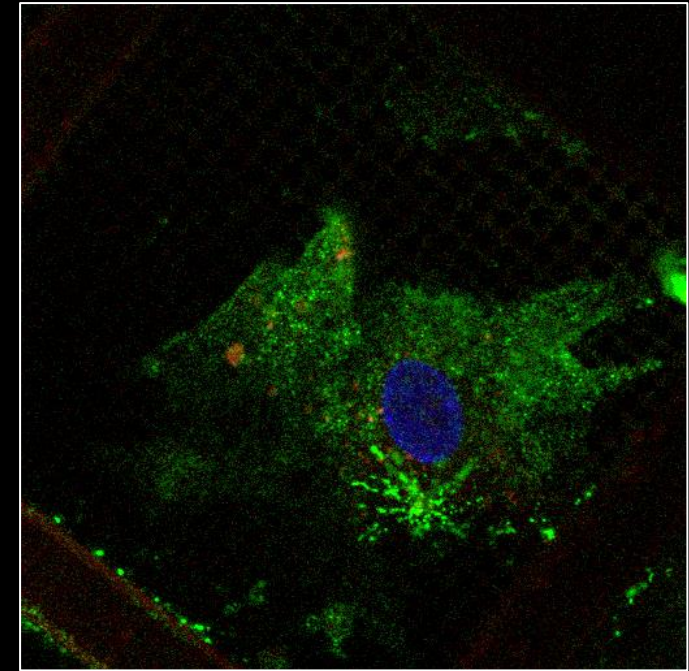


FURTHER STUDY:

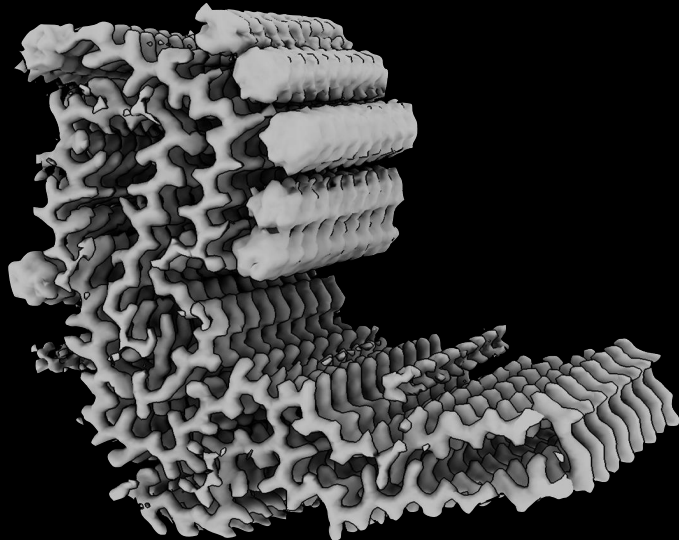
Cryo-FIB-ET



Prion infected astrocytes



RML



cryo-SXT/CLEXM



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Prion Unit

Prion strains

Adam Wenborn
Sue Joiner
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Prion cell biology

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Aline Marinho
Mitali Patel
Juan Ribes
Peter Kloehn
Parmjit Jat

MRC Prion Unit, Director

John Collinge



Supervision
Szymon Manka

Tilt-series alignment



Reconstructed tomogram

